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| (21) International Application Number: PCT/US99/06046 (22) International Filing Date: 19 March 1999 (19.03.99) (30) Priority Data: 60/079,386 26 March 1998 (26.03.98) US (71) Applicant (for all designated States except US): E.I. DU PONT DE NEMOURS AND COMPANY [US/US]; 1007 Market Street, Wilmington, DE 19898 (US). (72) Inventors; and (75) Inventors/Applicants (for US only): VOLLMER, Steven, J. [US/US]; 1019 Tweedbrook Road, Wilmington, DE 19810 (US). FALCO, Saverio, Carl [US/US]; 1902 Millers Road, Arden, DE 19810 (US). BROGLIE, Richard, M. [US/US]; 520 Port Royal Court, Landenberg, PA 19350 (US). BRYAN, Gregory, T. [NZ/US]; 1215 Spruce Street, Wilmington, DE 19805 (US). CAHOON, Rebecca, E. [US/US]; 2331 West 18th Street, Wilmington, DE 19806 (US). RAFALSKI, J., Antoni [US/US]; 2028 Longcome Drive, Wilmington, DE 19810 (US). (74) Agent: MAJARIAN, William, R.; E.I. du Pont de Nemours and Company, Legal Patent Records Center, 1007 Market Street, Wilmington, DE 19898 (US). | | (81) Designated States: AL, AU, BA, BB, BG, BR, CA, CN, CU, CZ, EE, GD, GE, HR, HU, ID, IL, IN, IS, JP, KP, KR, LC, LK, LR, LT, LV, MG, MK, MN, MX, NO, NZ, PL, RO, SG, SI, SK, SL, TR, TT, UA, US, UZ, VN, YU, ZA, ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG). Published <i>Without international search report and to be republished upon receipt of that report.</i> |
| (54) Title: TRYPTOPHAN BIOSYNTHETIC ENZYMES (57) Abstract This invention relates to an isolated nucleic acid fragment encoding a tryptophan biosynthetic enzyme. The invention also relates to the construction of a chimeric gene encoding all or a portion of the tryptophan biosynthetic enzyme, in sense or antisense orientation, wherein expression of the chimeric gene results in production of altered levels of the tryptophan biosynthetic enzyme in a transformed host cell. | | |

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TITLE

TRYPTOPHAN BIOSYNTHETIC ENZYMES

This application claims the benefit of U.S. Provisional Application No. 60/079,386, filed March 26, 1998.

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FIELD OF THE INVENTION

This invention is in the field of plant molecular biology. More specifically, this invention pertains to nucleic acid fragments encoding enzymes involved in the biosynthesis of tyrotophan in plants and seeds.

BACKGROUND OF THE INVENTION

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Many vertebrates, including man, lack the ability to manufacture a number of amino acids and therefore require these amino acids preformed in their diet. These are called essential amino acids. Plants are able to synthesize all twenty amino acids and serve as the ultimate source of the essential amino acids for humans and animals. Thus, the ability to manipulate the production and accumulation of the essential amino acids in plants is of considerable importance and value. Furthermore, the inability of animals to synthesize these amino acids provides a useful distinction between animal and plant cellular metabolism. This can be exploited for the discovery of herbicidal chemical compounds that target enzymes in the plant biosynthetic pathways of the essential amino acids and thus have low toxicity to animals.

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Tryptophan is an essential amino acid. In plants, the biosynthesis of tryptophan from chorismic acid (see Figure 1) requires five enzymatic steps catalyzed by anthranilate synthase (EC 4.1.3.27), anthranilate phosphoribosyl-transferase (EC 2.4.2.18), phosphoribosylanthranilate isomerase (EC 5.3.1.24), indole-3-glycerol phosphate synthase (EC 4.1.1.48) and tryptophan synthase (EC 4.2.1.20). The tryptophan pathway leads to the biosynthesis of many secondary metabolites including the hormone indole-3-acetic acid, antimicrobial phytoalexins, alkaloids and glucosinolates. The first enzyme in the tryptophan pathway from chorismate is anthranilate synthase. This enzyme is subject to feedback inhibition by tryptophan and is composed of two subunits. Even though the anthranilate synthase alpha subunits are encoded by duplicate genes, in *Arabidopsis* and *Ruta graveolens* it has been shown that only one gene is induced in response to wounding or pathogen attack (Bohlmann, J. et al. (1995) *Plant J.* 7:491-501 and Niyogi, K. K. and Fink, G. R. (1992) *Plant Cell* 4:721-733). Glutamine- and ammonia-dependent anthranilate synthase activities copurify in *Ruta graveolens*. The glutamine-dependent reaction of one of the alpha subunits requires a 60 to 65 kDa anthranilate synthase beta subunit (Bohlmann, J. et al. (1995) *Plant J.* 7:491-501) The beta-subunit is a highly asymmetric dimer with an apparent molecular weight of 200,000. Cleavage of the purified subunit with elastase, trypsin, or chymotrypsin results in fragments which retain enzyme activity. Elastase digestion results in a 30 kDa fragment and a 56 kDa fragment. The first fragment behaves as a monomer and interacts with free alpha subunit to produce the glutamine-dependent anthranilate synthase activity.

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The second fragment behaves as an asymmetric dimer and has N-(5'-phosphoribosyl) anthranilate isomerase and indole-3-glycerol phosphate synthase activity (Walker, M. S. and DeMoss, J. A. (1983) *J Biol Chem* 258:3571-3575).

5 The two final reactions in tryptophan biosynthesis are catalyzed by tryptophan synthase. The 29 kDa alpha subunit is a bifunctional enzyme which cleaves indole-3-glycerol phosphate to produce indole and glyceraldehyde-3-phosphate. The beta subunit joins indole with serine to form tryptophan. Either subunit alone is enzymatically active, but the rate of the reaction and affinity for the substrates increases when the subunits are forming a tetramer composed of two alpha subunits and two beta subunits (Radwanski, E. R. (1995) *Mol Gen Genet* 248:657-667).

Few of the genes encoding enzymes from the tryptophan pathway in corn, soybeans, rice and wheat, have been isolated and sequenced. For example, no corn, soybean, rice or wheat genes have been reported for anthranilate synthase alpha or beta subunits and no soybean, rice or wheat genes have been reported for tryptophan synthase alpha subunit. A
15 corn gene encoding tryptophan synthase has been identified and the instant invention describes a new corn tryptophan synthase homolog. Accordingly, the availability of nucleic acid sequences encoding all or a portion of these enzymes will facilitate studies to better understand cellular biosynthetic pathways, provide genetic tools for the manipulation of those pathways, provide a means to evaluate chemical compounds for their ability to inhibit
20 the enzymatic activity of the compounds included in this application.

SUMMARY OF THE INVENTION

The instant invention relates to isolated nucleic acid fragments encoding tryptophan biosynthetic enzymes. Specifically, this invention concerns an isolated nucleic acid fragment encoding an anthranilate synthase alpha subunit, an anthranilate synthase beta
25 subunit or a tryptophan synthase alpha subunit. In addition, this invention relates to a nucleic acid fragment that is complementary to the nucleic acid fragment encoding anthranilate synthase alpha subunit, anthranilate synthase beta subunit or tryptophan synthase alpha subunit.

An additional embodiment of the instant invention pertains to a polypeptide encoding
30 all or a substantial portion of a tryptophan biosynthetic enzyme selected from the group consisting of anthranilate synthase alpha subunit, anthranilate synthase beta subunit and tryptophan synthase alpha subunit.

In another embodiment, the instant invention relates to a chimeric gene encoding an anthranilate synthase alpha subunit, an anthranilate synthase beta subunit or a tryptophan
35 synthase alpha subunit, or to a chimeric gene that comprises a nucleic acid fragment that is complementary to a nucleic acid fragment encoding an anthranilate synthase alpha subunit, an anthranilate synthase beta subunit or a tryptophan synthase alpha subunit, operably linked to suitable regulatory sequences, wherein expression of the chimeric gene results in

production of levels of the encoded protein in a transformed host cell that is altered (i.e., increased or decreased) from the level produced in an untransformed host cell.

In a further embodiment, the instant invention concerns a transformed host cell comprising in its genome a chimeric gene encoding an anthranilate synthase alpha subunit, an anthranilate synthase beta subunit or a tryptophan synthase alpha subunit, operably linked to suitable regulatory sequences. Expression of the chimeric gene results in production of altered levels of the encoded protein in the transformed host cell. The transformed host cell can be of eukaryotic or prokaryotic origin, and include cells derived from higher plants and microorganisms. The invention also includes transformed plants that arise from transformed host cells of higher plants, and seeds derived from such transformed plants.

An additional embodiment of the instant invention concerns a method of altering the level of expression of an anthranilate synthase alpha subunit, an anthranilate synthase beta subunit or a tryptophan synthase alpha subunit in a transformed host cell comprising:
a) transforming a host cell with a chimeric gene comprising a nucleic acid fragment encoding an anthranilate synthase alpha subunit, an anthranilate synthase beta subunit or a tryptophan synthase alpha subunit; and b) growing the transformed host cell under conditions that are suitable for expression of the chimeric gene wherein expression of the chimeric gene results in production of altered levels of anthranilate synthase alpha subunit, anthranilate synthase beta subunit or tryptophan synthase alpha subunit in the transformed host cell.

An addition embodiment of the instant invention concerns a method for obtaining a nucleic acid fragment encoding all or a substantial portion of an amino acid sequence encoding an anthranilate synthase alpha subunit, an anthranilate synthase beta subunit or a tryptophan synthase alpha subunit.

A further embodiment of the instant invention is a method for evaluating at least one compound for its ability to inhibit the activity of an anthranilate synthase alpha subunit, an anthranilate synthase beta subunit or a tryptophan synthase alpha subunit, the method comprising the steps of: (a) transforming a host cell with a chimeric gene comprising a nucleic acid fragment encoding an anthranilate synthase alpha subunit, an anthranilate synthase beta subunit or a tryptophan synthase alpha subunit, operably linked to suitable regulatory sequences; (b) growing the transformed host cell under conditions that are suitable for expression of the chimeric gene wherein expression of the chimeric gene results in production of anthranilate synthase alpha subunit, anthranilate synthase beta subunit or tryptophan synthase alpha subunit in the transformed host cell; (c) optionally purifying the anthranilate synthase alpha subunit, the anthranilate synthase beta subunit or the tryptophan synthase alpha subunit expressed by the transformed host cell; (d) treating the anthranilate synthase alpha subunit, the anthranilate synthase beta subunit or the tryptophan synthase alpha subunit with a compound to be tested; and (e) comparing the activity of the anthranilate synthase alpha subunit, the anthranilate synthase beta subunit or the tryptophan

synthase alpha subunit that has been treated with a test compound to the activity of an untreated anthranilate synthase alpha subunit, anthranilate synthase beta subunit or tryptophan synthase alpha subunit, thereby selecting compounds with potential for inhibitory activity.

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BRIEF DESCRIPTION OF THE DRAWINGS AND SEQUENCE DESCRIPTIONS

The invention can be more fully understood from the following detailed description and the accompanying drawings and Sequence Listing which form a part of this application.

Figure 1 depicts biochemical pathway for the production of tryptophan from
10 chorismate.

Figure 2 shows an alignment of the amino acid sequences from *Ruta graveolens* anthranilate synthase alpha subunit (SEQ ID NO:39) and the instant corn anthranilate synthase alpha subunit (contig of cde1c.pk004.g2, p0125.czabf83r, p0005.cbmew77r, p0096.cnamt32r, p0032.crcas26r, p0031.ccmr07r, p0015.cdpfk12r, p0128.cpihl07r and
15 ctaln.pk0051.g3; SEQ ID NO:2). Amino acid which are identical among both sequences are indicated with an asterisk (*) above the alignment. Dashes are used by the program to maximize alignment of the sequences.

Figure 3 shows an alignment of the amino acid sequences from *Arabidopsis thaliana* anthranilate synthase beta subunit (SEQ ID NO:40), the instant corn anthranilate synthase
20 beta subunit (contig of p0126.cnlcu75r, cen3n.pk0212.h6, cco1n.pk0038.b1, csi1n.pk0017.d4, p0031.ccmr08r and ctn1c.pk001.110; SEQ ID NO:20) and the instant rice anthranilate synthase beta subunit (contig of rds2c.pk004.i9, rls24.pk0002.e2 and rl0n.pk113.b4; SEQ ID NO:22). Amino acid which are identical among all sequences are indicated with an asterisk (*) above the alignment. Dashes are used by the program to
25 maximize alignment of the sequences.

Figure 4 shows an alignment of the amino acid sequences from *Arabidopsis thaliana* tryptophan synthase alpha subunit (SEQ ID NO:41), the instant corn tryptophan synthase alpha subunit (contig of chp2.pk0020.e2 and lkr.pk0013.g1; SEQ ID NO:26), the instant
30 corn tryptophan synthase alpha subunit (cr1n.pk0052.b8; SEQ ID NO:28) and the instant rice tryptophan synthase alpha subunit (rr1.pk0038.h6; SEQ ID NO:32). Amino acid which are identical among all sequences are indicated with an asterisk (*) above the alignment. Dashes are used by the program to maximize alignment of the sequences.

The following sequence descriptions and Sequence Listing attached hereto comply with the rules governing nucleotide and/or amino acid sequence disclosures in patent
35 applications as set forth in 37 C.F.R. §1.821-1.825.

SEQ ID NO:1 is the nucleotide sequence comprising the contig assembled from a portion of the cDNA inserts in clones cde1c.pk004.g2, p0125.czabf83r, p0005.cbmew77r, p0096.cnamt32r, p0032.crcas26r, p0031.ccmr07r, p0015.cdpfk12r and p0128.cpihl07r and

the entire cDNA insert in clone cta1n.pk0051.g3 encoding an entire corn anthranilate synthase alpha subunit.

SEQ ID NO:2 is the deduced amino acid sequence of an entire corn anthranilate synthase alpha subunit derived from the nucleotide sequence of SEQ ID NO:1.

5 SEQ ID NO:3 is the nucleotide sequence comprising the contig assembled from a portion of the cDNA inserts in clones p0115.clsmj70r, cbn2.pk0049.d2, cpl1c.pk008.m4 and p0102.ceraq01r and the entire cDNA insert in clone ceb3.pk0011.c12 encoding a portion of a corn anthranilate synthase alpha subunit.

10 SEQ ID NO:4 is the deduced amino acid sequence of a portion of a corn anthranilate synthase alpha subunit derived from the nucleotide sequence of SEQ ID NO:3.

SEQ ID NO:5 is the nucleotide sequence comprising a portion of the cDNA insert in clone rca1n.pk004.p22 encoding a portion of a rice anthranilate synthase alpha subunit.

SEQ ID NO:6 is the deduced amino acid sequence of a portion of a rice anthranilate synthase alpha subunit derived from the nucleotide sequence of SEQ ID NO:5.

15 SEQ ID NO:7 is the nucleotide sequence comprising a portion of the cDNA insert in clone sdp3c.pk019.c6 encoding a portion of a soybean anthranilate synthase alpha subunit.

SEQ ID NO:8 is the deduced amino acid sequence of a portion of a soybean anthranilate synthase alpha subunit derived from the nucleotide sequence of SEQ ID NO:7.

20 SEQ ID NO:9 is the nucleotide sequence comprising a portion of the cDNA insert in clone sfl1.pk129.d10 encoding a portion of a soybean anthranilate synthase alpha subunit.

SEQ ID NO:10 is the deduced amino acid sequence of a portion of a soybean anthranilate synthase alpha subunit derived from the nucleotide sequence of SEQ ID NO:9.

25 SEQ ID NO:11 is the nucleotide sequence comprising a portion of the 5' terminus of the cDNA insert in clone wle1n.pk0075.b4 encoding a portion of a wheat anthranilate synthase alpha subunit.

SEQ ID NO:12 is the deduced amino acid sequence of a portion of a wheat anthranilate synthase alpha subunit derived from the nucleotide sequence of SEQ ID NO:11.

30 SEQ ID NO:13 is the nucleotide sequence comprising a portion of the 3' terminus of the cDNA insert in clone wle1n.pk0075.b4 encoding a portion of a wheat anthranilate synthase alpha subunit.

SEQ ID NO:14 is the deduced amino acid sequence of a portion of a wheat anthranilate synthase alpha subunit derived from the nucleotide sequence of SEQ ID NO:13.

SEQ ID NO:15 is the nucleotide sequence comprising a portion of the cDNA insert in clone wlm96.pk026.j1 encoding a portion of a wheat anthranilate synthase alpha subunit.

35 SEQ ID NO:16 is the deduced amino acid sequence of a portion of a wheat anthranilate synthase alpha subunit derived from the nucleotide sequence of SEQ ID NO:15.

SEQ ID NO:17 is the nucleotide sequence comprising the contig assembled from a portion of the cDNA insert in clones cen3n.pk0210.c3, cepe7.pk0012.g2 and cen3n.pk0047.g5 encoding a portion of a corn anthranilate synthase beta subunit.

SEQ ID NO:18 is the deduced amino acid sequence of a portion of a corn anthranilate synthase beta subunit derived from the nucleotide sequence of SEQ ID NO:17.

SEQ ID NO:19 is the nucleotide sequence comprising the contig assembled from a portion of the cDNA inserts in clones p0126.cnlcu75r, cen3n.pk0212.h6, cco1n.pk0038.b1, csiln.pk0017.d4, p0031.ccmal08r and ctn1c.pk001.110 encoding an entire corn anthranilate synthase beta subunit.

SEQ ID NO:20 is the deduced amino acid sequence of an entire corn anthranilate synthase beta subunit derived from the nucleotide sequence of SEQ ID NO:19.

SEQ ID NO:21 is the nucleotide sequence comprising the contig assembled from a portion of the cDNA inserts in clones rds2c.pk004.i9, rls24.pk0002.e2 and r10n.pk113.b4 encoding a substantial portion of a rice anthranilate synthase beta subunit.

SEQ ID NO:22 is the deduced amino acid sequence of a substantial portion of a rice anthranilate synthase beta subunit derived from the nucleotide sequence of SEQ ID NO:21.

SEQ ID NO:23 is the nucleotide sequence comprising a portion of the cDNA insert in clone wlm96.pk045.c11 encoding a portion of a wheat anthranilate synthase beta subunit.

SEQ ID NO:24 is the deduced amino acid sequence of a portion of a wheat anthranilate synthase beta subunit derived from the nucleotide sequence of SEQ ID NO:23.

SEQ ID NO:25 is the nucleotide sequence comprising the contig assembled from a portion of the cDNA inserts in clones chp2.pk0020.e2 and lkr.pk0013.g1 encoding an entire corn tryptophan synthase alpha subunit.

SEQ ID NO:26 is the deduced amino acid sequence of an entire corn tryptophan synthase alpha subunit derived from the nucleotide sequence of SEQ ID NO:25.

SEQ ID NO:27 is the nucleotide sequence comprising a portion of the cDNA insert in clone cr1n.pk0052.b8 encoding an almost entire corn tryptophan synthase alpha subunit.

SEQ ID NO:28 is the deduced amino acid sequence of an almost entire corn tryptophan synthase alpha subunit derived from the nucleotide sequence of SEQ ID NO:27.

SEQ ID NO:29 is the nucleotide sequence comprising the contig assembled from a portion of the cDNA inserts in clones res1c.pk004.c4 and rr1.pk0071.f2 encoding a portion of a rice tryptophan synthase alpha subunit.

SEQ ID NO:30 is the deduced amino acid sequence of a portion of a rice tryptophan synthase alpha subunit derived from the nucleotide sequence of SEQ ID NO:29.

SEQ ID NO:31 is the nucleotide sequence comprising a portion of the cDNA insert in clone rr1.pk0038.h6 encoding an entire rice tryptophan synthase alpha subunit.

SEQ ID NO:32 is the deduced amino acid sequence of an entire rice tryptophan synthase alpha subunit derived from the nucleotide sequence of SEQ ID NO:31.

SEQ ID NO:33 is the nucleotide sequence comprising the contig assembled from a portion of the cDNA inserts in clones src3c.pk004.c19 and sls2c.pk025.d11 encoding a portion of a soybean tryptophan synthase alpha subunit.

SEQ ID NO:34 is the deduced amino acid sequence of a portion of a soybean tryptophan synthase alpha subunit derived from the nucleotide sequence of SEQ ID NO:33.

SEQ ID NO:35 is the nucleotide sequence comprising the contig assembled from a portion of the cDNA inserts in clones wlm0.pk0028.f4 and wlm96.pk041.h3 encoding a portion of a wheat tryptophan synthase alpha subunit.

SEQ ID NO:36 is the deduced amino acid sequence of a portion of a wheat tryptophan synthase alpha subunit derived from the nucleotide sequence of SEQ ID NO:35.

SEQ ID NO:37 is the nucleotide sequence comprising the contig assembled from a portion of the cDNA inserts in clones wl1n.pk0109.d5 and wl1n.pk0110.c1 encoding a portion of a wheat tryptophan synthase alpha subunit.

SEQ ID NO:38 is the deduced amino acid sequence of a portion of a wheat tryptophan synthase alpha subunit derived from the nucleotide sequence of SEQ ID NO:37.

SEQ ID NO:39 is the amino acid sequence of a *Ruta graveolens* anthranilate synthase alpha subunit having an NCBI General Identifier No. 960289.

SEQ ID NO:40 is the amino acid sequence of a *Arabidopsis thaliana* anthranilate synthase beta subunit having an NCBI General Identifier No. 541849.

SEQ ID NO:41 is the amino acid sequence of a *Arabidopsis thaliana* tryptophan synthase alpha subunit having an NCBI General Identifier No. 2129755.

The Sequence Listing contains the one letter code for nucleotide sequence characters and the three letter codes for amino acids as defined in conformity with the IUPAC-IUBMB standards described in *Nucleic Acids Research* 13:3021-3030 (1985) and in the *Biochemical Journal* 219 (No. 2):345-373 (1984) which are herein incorporated by reference. The symbols and format used for nucleotide and amino acid sequence data comply with the rules set forth in 37 C.F.R. §1.822.

DETAILED DESCRIPTION OF THE INVENTION

In the context of this disclosure, a number of terms shall be utilized. As used herein, an "isolated nucleic acid fragment" is a polymer of RNA or DNA that is single- or double-stranded, optionally containing synthetic, non-natural or altered nucleotide bases. An isolated nucleic acid fragment in the form of a polymer of DNA may be comprised of one or more segments of cDNA, genomic DNA or synthetic DNA. As used herein, "contig" refers to an assemblage of overlapping nucleic acid sequences to form one contiguous nucleotide sequence. For example, several DNA sequences can be compared and aligned to identify common or overlapping regions. The individual sequences can then be assembled into a single contiguous nucleotide sequence.

As used herein, "substantially similar" refers to nucleic acid fragments wherein changes in one or more nucleotide bases results in substitution of one or more amino acids, but do not affect the functional properties of the protein encoded by the DNA sequence. "Substantially similar" also refers to nucleic acid fragments wherein changes in one or more nucleotide bases does not affect the ability of the nucleic acid fragment to mediate alteration

of gene expression by antisense or co-suppression technology. "Substantially similar" also refers to modifications of the nucleic acid fragments of the instant invention such as deletion or insertion of one or more nucleotides that do not substantially affect the functional properties of the resulting transcript vis-à-vis the ability to mediate alteration of gene expression by antisense or co-suppression technology or alteration of the functional properties of the resulting protein molecule. It is therefore understood that the invention encompasses more than the specific exemplary sequences.

For example, it is well known in the art that antisense suppression and co-suppression of gene expression may be accomplished using nucleic acid fragments representing less than the entire coding region of a gene, and by nucleic acid fragments that do not share 100% sequence identity with the gene to be suppressed. Moreover, alterations in a gene which result in the production of a chemically equivalent amino acid at a given site, but do not effect the functional properties of the encoded protein, are well known in the art. Thus, a codon for the amino acid alanine, a hydrophobic amino acid, may be substituted by a codon encoding another less hydrophobic residue, such as glycine, or a more hydrophobic residue, such as valine, leucine, or isoleucine. Similarly, changes which result in substitution of one negatively charged residue for another, such as aspartic acid for glutamic acid, or one positively charged residue for another, such as lysine for arginine, can also be expected to produce a functionally equivalent product. Nucleotide changes which result in alteration of the N-terminal and C-terminal portions of the protein molecule would also not be expected to alter the activity of the protein. Each of the proposed modifications is well within the routine skill in the art, as is determination of retention of biological activity of the encoded products. Moreover, the skilled artisan recognizes that substantially similar nucleic acid sequences encompassed by this invention are also defined by their ability to hybridize, under stringent conditions (0.1X SSC, 0.1% SDS, 65°C), with the sequences exemplified herein. Substantially similar nucleic acid fragments of the instant invention may also be characterized by the percent identity of the amino acid sequences that they encode to the amino acid sequences disclosed herein, as determined by algorithms commonly employed by those skilled in this art. Preferred are those nucleic acid fragments whose nucleotide sequences encode amino acid sequences that are greater than 80% identical to the amino acid sequences reported herein. More preferred nucleic acid fragments encode amino acid sequences that are 90% identical to the amino acid sequences reported herein. Most preferred are nucleic acid fragments that encode amino acid sequences that are 95% identical to the amino acid sequences reported herein. Sequence alignments and percent similarity calculations were performed using the Megalign program of the LASARGENE bioinformatics computing suite (DNASTAR Inc., Madison, WI). Multiple alignment of the sequences was performed using the Clustal method of alignment (Higgins, D.G. and Sharp, P.M. (1989) *CABIOS*. 5:151-153) with the default parameters (GAP PENALTY=10, GAP

LENGTH PENALTY=10). Default parameters for pairwise alignments using the Clustal method were KTUPLE 1, GAP PENALTY=3, WINDOW=5 and DIAGONALS SAVED=5.

A “substantial portion” of an amino acid or nucleotide sequence comprises enough of the amino acid sequence of a polypeptide or the nucleotide sequence of a gene to afford putative identification of that polypeptide or gene, either by manual evaluation of the sequence by one skilled in the art, or by computer-automated sequence comparison and identification using algorithms such as BLAST (Basic Local Alignment Search Tool; Altschul, S. F., et al., (1993) *J. Mol. Biol.* 215:403-410; see also www.ncbi.nlm.nih.gov/BLAST/). In general, a sequence of ten or more contiguous amino acids or thirty or more nucleotides is necessary in order to putatively identify a polypeptide or nucleic acid sequence as homologous to a known protein or gene. Moreover, with respect to nucleotide sequences, gene specific oligonucleotide probes comprising 20-30 contiguous nucleotides may be used in sequence-dependent methods of gene identification (e.g., Southern hybridization) and isolation (e.g., *in situ* hybridization of bacterial colonies or bacteriophage plaques). In addition, short oligonucleotides of 12-15 bases may be used as amplification primers in PCR in order to obtain a particular nucleic acid fragment comprising the primers. Accordingly, a “substantial portion” of a nucleotide sequence comprises enough of the sequence to afford specific identification and/or isolation of a nucleic acid fragment comprising the sequence. The instant specification teaches partial or complete amino acid and nucleotide sequences encoding one or more particular plant proteins. The skilled artisan, having the benefit of the sequences as reported herein, may now use all or a substantial portion of the disclosed sequences for purposes known to those skilled in this art. Accordingly, the instant invention comprises the complete sequences as reported in the accompanying Sequence Listing, as well as substantial portions of those sequences as defined above.

“Codon degeneracy” refers to divergence in the genetic code permitting variation of the nucleotide sequence without effecting the amino acid sequence of an encoded polypeptide. Accordingly, the instant invention relates to any nucleic acid fragment that encodes all or a substantial portion of the amino acid sequence encoding the anthranilate synthase alpha subunit, the anthranilate synthase beta subunit or the tryptophan synthase alpha subunit proteins as set forth in SEQ ID NOs:2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36 and 38. The skilled artisan is well aware of the “codon-bias” exhibited by a specific host cell in usage of nucleotide codons to specify a given amino acid. Therefore, when synthesizing a gene for improved expression in a host cell, it is desirable to design the gene such that its frequency of codon usage approaches the frequency of preferred codon usage of the host cell.

“Synthetic genes” can be assembled from oligonucleotide building blocks that are chemically synthesized using procedures known to those skilled in the art. These building blocks are ligated and annealed to form gene segments which are then enzymatically

- assembled to construct the entire gene. "Chemically synthesized", as related to a sequence of DNA, means that the component nucleotides were assembled *in vitro*. Manual chemical synthesis of DNA may be accomplished using well established procedures, or automated chemical synthesis can be performed using one of a number of commercially available machines. Accordingly, the genes can be tailored for optimal gene expression based on optimization of nucleotide sequence to reflect the codon bias of the host cell. The skilled artisan appreciates the likelihood of successful gene expression if codon usage is biased towards those codons favored by the host. Determination of preferred codons can be based on a survey of genes derived from the host cell where sequence information is available.
- 10 "Gene" refers to a nucleic acid fragment that expresses a specific protein, including regulatory sequences preceding (5' non-coding sequences) and following (3' non-coding sequences) the coding sequence. "Native gene" refers to a gene as found in nature with its own regulatory sequences. "Chimeric gene" refers any gene that is not a native gene, comprising regulatory and coding sequences that are not found together in nature.
- 15 Accordingly, a chimeric gene may comprise regulatory sequences and coding sequences that are derived from different sources, or regulatory sequences and coding sequences derived from the same source, but arranged in a manner different than that found in nature. "Endogenous gene" refers to a native gene in its natural location in the genome of an organism. A "foreign" gene refers to a gene not normally found in the host organism, but
- 20 that is introduced into the host organism by gene transfer. Foreign genes can comprise native genes inserted into a non-native organism, or chimeric genes. A "transgene" is a gene that has been introduced into the genome by a transformation procedure.
- "Coding sequence" refers to a DNA sequence that codes for a specific amino acid sequence. "Regulatory sequences" refer to nucleotide sequences located upstream (5' non-coding sequences), within, or downstream (3' non-coding sequences) of a coding sequence, and which influence the transcription, RNA processing or stability, or translation of the associated coding sequence. Regulatory sequences may include promoters, translation leader sequences, introns, and polyadenylation recognition sequences.
- 25 "Promoter" refers to a DNA sequence capable of controlling the expression of a coding sequence or functional RNA. In general, a coding sequence is located 3' to a promoter sequence. The promoter sequence consists of proximal and more distal upstream elements, the latter elements often referred to as enhancers. Accordingly, an "enhancer" is a DNA sequence which can stimulate promoter activity and may be an innate element of the promoter or a heterologous element inserted to enhance the level or tissue-specificity of a
- 30 promoter. Promoters may be derived in their entirety from a native gene, or be composed of different elements derived from different promoters found in nature, or even comprise synthetic DNA segments. It is understood by those skilled in the art that different promoters may direct the expression of a gene in different tissues or cell types, or at different stages of development, or in response to different environmental conditions. Promoters which cause a

gene to be expressed in most cell types at most times are commonly referred to as “constitutive promoters”. New promoters of various types useful in plant cells are constantly being discovered; numerous examples may be found in the compilation by Okamuro and Goldberg, (1989) *Biochemistry of Plants* 15:1-82. It is further recognized that since in most cases the exact boundaries of regulatory sequences have not been completely defined, DNA fragments of different lengths may have identical promoter activity.

The “translation leader sequence” refers to a DNA sequence located between the promoter sequence of a gene and the coding sequence. The translation leader sequence is present in the fully processed mRNA upstream of the translation start sequence. The translation leader sequence may affect processing of the primary transcript to mRNA, mRNA stability or translation efficiency. Examples of translation leader sequences have been described (Turner, R. and Foster, G. D. (1995) *Molecular Biotechnology* 3:225).

The “3' non-coding sequences” refer to DNA sequences located downstream of a coding sequence and include polyadenylation recognition sequences and other sequences encoding regulatory signals capable of affecting mRNA processing or gene expression. The polyadenylation signal is usually characterized by affecting the addition of polyadenylic acid tracts to the 3' end of the mRNA precursor. The use of different 3' non-coding sequences is exemplified by Ingelbrecht et al., (1989) *Plant Cell* 1:671-680.

“RNA transcript” refers to the product resulting from RNA polymerase-catalyzed transcription of a DNA sequence. When the RNA transcript is a perfect complementary copy of the DNA sequence, it is referred to as the primary transcript or it may be a RNA sequence derived from posttranscriptional processing of the primary transcript and is referred to as the mature RNA. “Messenger RNA (mRNA)” refers to the RNA that is without introns and that can be translated into protein by the cell. “cDNA” refers to a double-stranded DNA that is complementary to and derived from mRNA. “Sense” RNA refers to RNA transcript that includes the mRNA and so can be translated into protein by the cell. “Antisense RNA” refers to a RNA transcript that is complementary to all or part of a target primary transcript or mRNA and that blocks the expression of a target gene (U.S. Pat. No. 5,107,065, incorporated herein by reference). The complementarity of an antisense RNA may be with any part of the specific gene transcript, i.e., at the 5' non-coding sequence, 3' non-coding sequence, introns, or the coding sequence. “Functional RNA” refers to sense RNA, antisense RNA, ribozyme RNA, or other RNA that may not be translated but yet has an effect on cellular processes.

The term “operably linked” refers to the association of nucleic acid sequences on a single nucleic acid fragment so that the function of one is affected by the other. For example, a promoter is operably linked with a coding sequence when it is capable of affecting the expression of that coding sequence (i.e., that the coding sequence is under the transcriptional control of the promoter). Coding sequences can be operably linked to regulatory sequences in sense or antisense orientation.

The term "expression", as used herein, refers to the transcription and stable accumulation of sense (mRNA) or antisense RNA derived from the nucleic acid fragment of the invention. Expression may also refer to translation of mRNA into a polypeptide.

"Antisense inhibition" refers to the production of antisense RNA transcripts capable of suppressing the expression of the target protein. "Overexpression" refers to the production of a gene product in transgenic organisms that exceeds levels of production in normal or non-transformed organisms. "Co-suppression" refers to the production of sense RNA transcripts capable of suppressing the expression of identical or substantially similar foreign or endogenous genes (U.S. Pat. No. 5,231,020, incorporated herein by reference).

"Altered levels" refers to the production of gene product(s) in transgenic organisms in amounts or proportions that differ from that of normal or non-transformed organisms.

"Mature" protein refers to a post-translationally processed polypeptide; i.e., one from which any pre- or propeptides present in the primary translation product have been removed. "Precursor" protein refers to the primary product of translation of mRNA; i.e., with pre- and propeptides still present. Pre- and propeptides may be but are not limited to intracellular localization signals.

A "chloroplast transit peptide" is an amino acid sequence which is translated in conjunction with a protein and directs the protein to the chloroplast or other plastid types present in the cell in which the protein is made. "Chloroplast transit sequence" refers to a nucleotide sequence that encodes a chloroplast transit peptide. A "signal peptide" is an amino acid sequence which is translated in conjunction with a protein and directs the protein to the secretory system (Chrispeels, J. J., (1991) *Ann. Rev. Plant Phys. Plant Mol. Biol.* 42:21-53). If the protein is to be directed to a vacuole, a vacuolar targeting signal (*supra*) can further be added, or if to the endoplasmic reticulum, an endoplasmic reticulum retention signal (*supra*) may be added. If the protein is to be directed to the nucleus, any signal peptide present should be removed and instead a nuclear localization signal included (Raikhel (1992) *Plant Phys.* 100:1627-1632).

"Transformation" refers to the transfer of a nucleic acid fragment into the genome of a host organism, resulting in genetically stable inheritance. Host organisms containing the transformed nucleic acid fragments are referred to as "transgenic" organisms. Examples of methods of plant transformation include *Agrobacterium*-mediated transformation (De Blaere et al. (1987) *Meth. Enzymol.* 143:277) and particle-accelerated or "gene gun" transformation technology (Klein et al. (1987) *Nature (London)* 327:70-73; U.S. Pat. No. 4,945,050, incorporated herein by reference).

Standard recombinant DNA and molecular cloning techniques used herein are well known in the art and are described more fully in Sambrook, J., Fritsch, E.F. and Maniatis, T. *Molecular Cloning: A Laboratory Manual*; Cold Spring Harbor Laboratory Press: Cold Spring Harbor, 1989 (hereinafter "Maniatis").

- Nucleic acid fragments encoding at least a portion of several tryptophan biosynthetic enzymes have been isolated and identified by comparison of random plant cDNA sequences to public databases containing nucleotide and protein sequences using the BLAST algorithms well known to those skilled in the art. Table 1 lists the proteins that are described herein, and the designation of the cDNA clones that comprise the nucleic acid fragments encoding these proteins.

TABLE 1

Tryptophan Biosynthetic Enzymes

| Enzyme | Clone | Plant |
|-------------------------------------|--|---------|
| Anthranilate Synthase Alpha Subunit | Contig of: cde1c.pk004.g2 p0125.czabf83r p0005.cbmew77r p0096.cnamt32r p0032.crcas26r p0031.ccmr07r p0015.cdpfk12r p0128.cpihl07r cta1n.pk0051.g3 | Corn |
| | Contig of: p0115.clsmj70r cbn2.pk0049.d2 cpl1c.pk008.m4 p0102.ceraq01r ceb3.pk0011.c12 | Corn |
| | rca1n.pk004.p22 | Rice |
| | sdp3c.pk019.c6 | Soybean |
| | sfl1.pk129.d10 | Soybean |
| | wle1n.pk0075.b4 | Wheat |
| | wlm96.pk026.j1 | Wheat |
| | Contig of: cen3n.pk0210.c3 cepe7.pk0012.g2 cen3n.pk0047.g5 | Corn |
| | Contig of: p0126.cnlcu75r cen3n.pk0212.h6 cco1n.pk0038.b1 csi1n.pk0017.d4 p0031.ccmr08r ctn1c.pk001.110 | Corn |
| | Contig of: rds2c.pk004.i9 rls24.pk0002.e2 rl0n.pk113.b4 | Rice |
| Anthranilate Synthase Beta Subunit | wlm96.pk045.c11 | Wheat |

| Enzyme | Clone | Plant |
|-----------------------------------|--|---------|
| Tryptophan Synthase Alpha Subunit | Contig of: chp2.pk0020.e2 lkr.pk0013.g1 | Corn |
| | cr1n.pk0052.b8 | Corn |
| | Contig of: res1c.pk004.c4 rr1.pk0071.f2 | Rice |
| | rr1.pk0038.h6 | Rice |
| | Contig of: src3c.pk004.c19 sls2c.pk025.d11 | Soybean |
| | Contig of: wlm0.pk0028.f4 wlm96.pk041.h3 | Wheat |
| | Contig of: wl1n.pk0109.d5 wl1n.pk0110.c1 | Wheat |
| | | |

- The nucleic acid fragments of the instant invention may be used to isolate cDNAs and genes encoding homologous proteins from the same or other plant species. Isolation of homologous genes using sequence-dependent protocols is well known in the art. Examples of sequence-dependent protocols include, but are not limited to, methods of nucleic acid hybridization, and methods of DNA and RNA amplification as exemplified by various uses of nucleic acid amplification technologies (e.g., polymerase chain reaction, ligase chain reaction).
- For example, genes encoding other anthranilate synthase alpha subunit, anthranilate synthase beta subunit or tryptophan synthase alpha subunit, either as cDNAs or genomic DNAs, could be isolated directly by using all or a portion of the instant nucleic acid fragments as DNA hybridization probes to screen libraries from any desired plant employing methodology well known to those skilled in the art. Specific oligonucleotide probes based upon the instant nucleic acid sequences can be designed and synthesized by methods known in the art (Maniatis). Moreover, the entire sequences can be used directly to synthesize DNA probes by methods known to the skilled artisan such as random primer DNA labeling, nick translation, or end-labeling techniques, or RNA probes using available *in vitro* transcription systems. In addition, specific primers can be designed and used to amplify a part or all of the instant sequences. The resulting amplification products can be labeled directly during amplification reactions or labeled after amplification reactions, and used as probes to isolate full length cDNA or genomic fragments under conditions of appropriate stringency.

In addition, two short segments of the instant nucleic acid fragments may be used in polymerase chain reaction protocols to amplify longer nucleic acid fragments encoding

homologous genes from DNA or RNA. The polymerase chain reaction may also be performed on a library of cloned nucleic acid fragments wherein the sequence of one primer is derived from the instant nucleic acid fragments, and the sequence of the other primer takes advantage of the presence of the polyadenylic acid tracts to the 3' end of the mRNA

5 precursor encoding plant genes. Alternatively, the second primer sequence may be based upon sequences derived from the cloning vector. For example, the skilled artisan can follow the RACE protocol (Frohman et al., (1988) *PNAS USA* 85:8998) to generate cDNAs by using PCR to amplify copies of the region between a single point in the transcript and the 3' or 5' end. Primers oriented in the 3' and 5' directions can be designed from the instant
10 sequences. Using commercially available 3' RACE or 5' RACE systems (BRL), specific 3' or 5' cDNA fragments can be isolated (Ohara et al., (1989) *PNAS USA* 86:5673; Loh et al., (1989) *Science* 243:217). Products generated by the 3' and 5' RACE procedures can be combined to generate full-length cDNAs (Frohman, M. A. and Martin, G. R., (1989) *Techniques* 1:165).

15 Availability of the instant nucleotide and deduced amino acid sequences facilitates immunological screening of cDNA expression libraries. Synthetic peptides representing portions of the instant amino acid sequences may be synthesized. These peptides can be used to immunize animals to produce polyclonal or monoclonal antibodies with specificity for peptides or proteins comprising the amino acid sequences. These antibodies can be then
20 be used to screen cDNA expression libraries to isolate full-length cDNA clones of interest (Lerner, R. A. (1984) *Adv. Immunol.* 36:1; Maniatis).

The nucleic acid fragments of the instant invention may be used to create transgenic plants in which the disclosed anthranilate synthase alpha subunit, anthranilate synthase beta subunit or tryptophan synthase alpha subunit are present at higher or lower levels than
25 normal or in cell types or developmental stages in which they are not normally found. This would have the effect of altering the level of tryptophan in those cells. Manipulation of the levels of some of the anthranilate synthase alpha subunits will also result in changes in the response to pathogen attack. Because this pathway is not followed for the production of tryptophan in higher animals, these enzymes are very good candidates for the discovery of
30 herbicides and fungicides.

Overexpression of the anthranilate synthase alpha subunit, the anthranilate synthase beta subunit or the tryptophan synthase alpha subunit proteins of the instant invention may be accomplished by first constructing a chimeric gene in which the coding region is operably linked to a promoter capable of directing expression of a gene in the desired tissues at the
35 desired stage of development. For reasons of convenience, the chimeric gene may comprise promoter sequences and translation leader sequences derived from the same genes. 3' Non-coding sequences encoding transcription termination signals may also be provided. The instant chimeric gene may also comprise one or more introns in order to facilitate gene expression.

Plasmid vectors comprising the instant chimeric gene can then constructed. The choice of plasmid vector is dependent upon the method that will be used to transform host plants. The skilled artisan is well aware of the genetic elements that must be present on the plasmid vector in order to successfully transform, select and propagate host cells containing the chimeric gene. The skilled artisan will also recognize that different independent transformation events will result in different levels and patterns of expression (Jones et al., (1985) *EMBO J.* 4:2411-2418; De Almeida et al., (1989) *Mol. Gen. Genetics* 218:78-86), and thus that multiple events must be screened in order to obtain lines displaying the desired expression level and pattern. Such screening may be accomplished by Southern analysis of DNA, Northern analysis of mRNA expression, Western analysis of protein expression, or phenotypic analysis.

For some applications it may be useful to direct the instant tryptophan biosynthetic enzyme to different cellular compartments, or to facilitate its secretion from the cell. It is thus envisioned that the chimeric gene described above may be further supplemented by altering the coding sequence to encode anthranilate synthase alpha subunit, anthranilate synthase beta subunit or tryptophan synthase alpha subunit with appropriate intracellular targeting sequences such as transit sequences (Keegstra, K. (1989) *Cell* 56:247-253), signal sequences or sequences encoding endoplasmic reticulum localization (Chrispeels, J.J., (1991) *Ann. Rev. Plant Phys. Plant Mol. Biol.* 42:21-53), or nuclear localization signals (Raikhel, N. (1992) *Plant Phys.* 100:1627-1632) added and/or with targeting sequences that are already present removed. While the references cited give examples of each of these, the list is not exhaustive and more targeting signals of utility may be discovered in the future.

It may also be desirable to reduce or eliminate expression of genes encoding anthranilate synthase alpha subunit, anthranilate synthase beta subunit or tryptophan synthase alpha subunit in plants for some applications. In order to accomplish this, a chimeric gene designed for co-suppression of the instant tryptophan biosynthetic enzyme can be constructed by linking a gene or gene fragment encoding an anthranilate synthase alpha subunit, anthranilate synthase beta subunit or tryptophan synthase alpha subunit to plant promoter sequences. Alternatively, a chimeric gene designed to express antisense RNA for all or part of the instant nucleic acid fragment can be constructed by linking the gene or gene fragment in reverse orientation to plant promoter sequences. Either the co-suppression or antisense chimeric genes could be introduced into plants via transformation wherein expression of the corresponding endogenous genes are reduced or eliminated.

The instant anthranilate synthase alpha subunit, anthranilate synthase beta subunit or tryptophan synthase alpha subunit (or portions thereof) may be produced in heterologous host cells, particularly in the cells of microbial hosts, and can be used to prepare antibodies to these proteins by methods well known to those skilled in the art. The antibodies are useful for detecting anthranilate synthase alpha subunit, anthranilate synthase beta subunit or tryptophan synthase alpha subunit *in situ* in cells or *in vitro* in cell extracts. Preferred

heterologous host cells for production of the instant anthranilate synthase alpha subunit, anthranilate synthase beta subunit or tryptophan synthase alpha subunit are microbial hosts. Microbial expression systems and expression vectors containing regulatory sequences that direct high level expression of foreign proteins are well known to those skilled in the art.

- 5 Any of these could be used to construct a chimeric gene for production of the instant anthranilate synthase alpha subunit, anthranilate synthase beta subunit or tryptophan synthase alpha subunit. This chimeric gene could then be introduced into appropriate microorganisms via transformation to provide high level expression of the encoded tryptophan biosynthetic enzyme. An example of a vector for high level expression of the
10 instant anthranilate synthase alpha subunit, anthranilate synthase beta subunit or tryptophan synthase alpha subunit in a bacterial host is provided (Example 7).

- Additionally, the instant anthranilate synthase alpha subunit, anthranilate synthase beta subunit or tryptophan synthase alpha subunit can be used as targets to facilitate design and/or identification of inhibitors of those enzymes that may be useful as herbicides. This is
15 desirable because the anthranilate synthase alpha subunit, the anthranilate synthase beta subunit and the tryptophan synthase alpha subunit described herein catalyze various steps in tryptophan biosynthesis. Accordingly, inhibition of the activity of one or more of the enzymes described herein could lead to inhibition plant growth. Thus, the instant anthranilate synthase alpha subunit, anthranilate synthase beta subunit, tryptophan synthase
20 alpha subunit could be appropriate for new herbicide discovery and design.

- All or a substantial portion of the nucleic acid fragments of the instant invention may also be used as probes for genetically and physically mapping the genes that they are a part of, and as markers for traits linked to those genes. Such information may be useful in plant breeding in order to develop lines with desired phenotypes. For example, the instant nucleic
25 acid fragments may be used as restriction fragment length polymorphism (RFLP) markers. Southern blots (Maniatis) of restriction-digested plant genomic DNA may be probed with the nucleic acid fragments of the instant invention. The resulting banding patterns may then be subjected to genetic analyses using computer programs such as MapMaker (Lander et al., (1987) *Genomics* 1:174-181) in order to construct a genetic map. In addition, the nucleic
30 acid fragments of the instant invention may be used to probe Southern blots containing restriction endonuclease-treated genomic DNAs of a set of individuals representing parent and progeny of a defined genetic cross. Segregation of the DNA polymorphisms is noted and used to calculate the position of the instant nucleic acid sequence in the genetic map previously obtained using this population (Botstein, D. et al., (1980) *Am. J. Hum. Genet.*
35 32:314-331).

The production and use of plant gene-derived probes for use in genetic mapping is described in R. Bernatzky, R. and Tanksley, S. D. (1986) *Plant Mol. Biol. Reporter* 4(1):37-41. Numerous publications describe genetic mapping of specific cDNA clones using the methodology outlined above or variations thereof. For example, F2 intercross

populations, backcross populations, randomly mated populations, near isogenic lines, and other sets of individuals may be used for mapping. Such methodologies are well known to those skilled in the art.

5 Nucleic acid probes derived from the instant nucleic acid sequences may also be used for physical mapping (i.e., placement of sequences on physical maps; *see* Hoheisel, J. D., et al., In: *Nonmammalian Genomic Analysis: A Practical Guide*, Academic press 1996, pp. 319-346, and references cited therein).

10 In another embodiment, nucleic acid probes derived from the instant nucleic acid sequences may be used in direct fluorescence *in situ* hybridization (FISH) mapping (Trask, B. J. (1991) *Trends Genet.* 7:149-154). Although current methods of FISH mapping favor use of large clones (several to several hundred KB; *see* Laan, M. et al. (1995) *Genome Research* 5:13-20), improvements in sensitivity may allow performance of FISH mapping using shorter probes.

15 A variety of nucleic acid amplification-based methods of genetic and physical mapping may be carried out using the instant nucleic acid sequences. Examples include allele-specific amplification (Kazazian, H. H. (1989) *J. Lab. Clin. Med.* 114(2):95-96), polymorphism of PCR-amplified fragments (CAPS; Sheffield, V. C. et al. (1993) *Genomics* 16:325-332), allele-specific ligation (Landegren, U. et al. (1988) *Science* 241:1077-1080), nucleotide extension reactions (Sokolov, B. P. (1990) *Nucleic Acid Res.* 18:3671), Radiation
20 Hybrid Mapping (Walter, M. A. et al. (1997) *Nature Genetics* 7:22-28) and Happy Mapping (Dear, P. H. and Cook, P. R. (1989) *Nucleic Acid Res.* 17:6795-6807). For these methods, the sequence of a nucleic acid fragment is used to design and produce primer pairs for use in the amplification reaction or in primer extension reactions. The design of such primers is well known to those skilled in the art. In methods employing PCR-based genetic mapping,
25 it may be necessary to identify DNA sequence differences between the parents of the mapping cross in the region corresponding to the instant nucleic acid sequence. This, however, is generally not necessary for mapping methods.

Loss of function mutant phenotypes may be identified for the instant cDNA clones either by targeted gene disruption protocols or by identifying specific mutants for these
30 genes contained in a maize population carrying mutations in all possible genes (Ballinger and Benzer, (1989) *Proc. Natl. Acad. Sci USA* 86:9402; Koes et al., (1995) *Proc. Natl. Acad. Sci USA* 92:8149; Bensen et al., (1995) *Plant Cell* 7:75). The latter approach may be accomplished in two ways. First, short segments of the instant nucleic acid fragments may be used in polymerase chain reaction protocols in conjunction with a mutation tag sequence
35 primer on DNAs prepared from a population of plants in which Mutator transposons or some other mutation-causing DNA element has been introduced (*see* Bensen, *supra*). The amplification of a specific DNA fragment with these primers indicates the insertion of the mutation tag element in or near the plant gene encoding the anthranilate synthase alpha subunit, the anthranilate synthase beta subunit or the tryptophan synthase alpha subunit.

Alternatively, the instant nucleic acid fragment may be used as a hybridization probe against PCR amplification products generated from the mutation population using the mutation tag sequence primer in conjunction with an arbitrary genomic site primer, such as that for a restriction enzyme site-anchored synthetic adaptor. With either method, a plant containing a mutation in the endogenous gene encoding an anthranilate synthase alpha subunit, an anthranilate synthase beta subunit or a tryptophan synthase alpha subunit can be identified and obtained. This mutant plant can then be used to determine or confirm the natural function of the anthranilate synthase alpha subunit, the anthranilate synthase beta subunit or the tryptophan synthase alpha subunit gene product.

EXAMPLES

The present invention is further defined in the following Examples, in which all parts and percentages are by weight and degrees are Celsius, unless otherwise stated. It should be understood that these Examples, while indicating preferred embodiments of the invention, are given by way of illustration only. From the above discussion and these Examples, one skilled in the art can ascertain the essential characteristics of this invention, and without departing from the spirit and scope thereof, can make various changes and modifications of the invention to adapt it to various usages and conditions.

EXAMPLE 1

Composition of cDNA Libraries; Isolation and Sequencing of cDNA Clones

cDNA libraries representing mRNAs from various corn, rice, soybean and wheat tissues were prepared. The characteristics of the libraries are described below.

TABLE 2

cDNA Libraries from Corn, Rice, Soybean and Wheat

| Library | Tissue | Clone |
|---------|---|---|
| cbn2 | Corn Developing Kernel Two Days After Pollination | cbn2.pk0049.d2 |
| cco1n | Corn Cob of 67 Day Old Plants Grown in Green House* | cco1n.pk0038.b1 |
| cde1c | Corn Developing Embryo 20 Days After Pollination | cde1c.pk004.g2 |
| ceb3 | Corn Embryo 20 Days After Pollination | ceb3.pk0011.c12 |
| cen3n | Corn Endosperm 20 Days After Pollination* | cen3n.pk0047.g5 cen3n.pk0210.c3 cen3n.pk0212.h6 |
| cepe7 | Corn 7 Day Old Epicotyl From Etiolated Seedling | cepe7.pk0012.g2 |
| chp2 | Corn (B73 and MK593) 11 Day Old Leaf Treated 24 Hours With Herbicides** | chp2.pk0020.e2 |
| cpl1c | Corn Pooled BMS Treated With Chemical Chelators*** | cpl1c.pk008.m4 |
| cr1n | Corn Root From 7 Day Old Seedlings* | cr1n.pk0052.b8 |

| Library | Tissue | Clone |
|---------|---|---|
| csiln | Corn Silk* | csiln.pk0017.d4 |
| ctaln | Corn Tassel* | ctaln.pk0051.g3 |
| ctn1c | Corn Tassel, Night Harvested | ctn1c.pk001.110 |
| lkr | Corn 19 Day Old Seed | lkr.pk0013.g1 |
| p0005 | Corn Immature Ear | p0005.cbmew77r |
| p0015 | Corn Embryo 13 Days After Pollination | p0015.cdpfk12r |
| p0031 | Corn (CM45) Shoot Culture | p0031.ccmmai08r p0031.ccmar07r |
| p0032 | Corn Regenerating Callus (Hi-II 223a and 1129e), 10 and 14 Days After Auxin Removal, Pooled | p0032.crcas26r |
| p0096 | Corn Scutellum 2 and 3 Days After Germinating | p0096.cnamt32r |
| p0102 | Corn Early Meiosis Tassels* | p0102.ceraq01r |
| p0115 | Corn Leaf and Sheath Meristem Tissue Collected from 10th, 11th, and 12th Leaves, Pooled | p0115.clsmj70r |
| p0125 | Corn Anther Prophase I | p0125.czabf83r |
| p0126 | Corn Leaf Tissue (V8-V10****), Night-Harvested | p0126.cnlcu75r |
| p0128 | Corn Primary and Secondary Immature Ear | p0128.cpihl07r |
| rca1n | Rice Callus* | rca1n.pk004.p22 |
| rds2c | Rice Developing Seeds | rds2c.pk004.i9 |
| res1c | Rice Etiolated Seedling | res1c.pk004.c4 |
| rl0n | Rice 15 Day Old Leaf* | rl0n.pk113.b4 |
| rls24 | Rice Leaf 15 Days After Germination, 24 Hours After Infection of Strain <i>Magaporthe grisea</i> 4360-R-67 (AVR2-YAMO); Susceptible | rls24.pk0002.e2 |
| rr1 | Rice Root of Two Week Old Developing Seedling | rr1.pk0038.h6 rr1.pk0071.f2 |
| sdp3c | Soybean Developing Pods (8-9 mm) | sdp3c.pk019.c6 |
| sfl1 | Soybean Immature Flower | sfl1.pk129.d10 |
| sls2c | Soybean Infected With <i>Sclerotinia sclerotiorum</i> mycelium | sls2c.pk025.d11 |
| src3c | Soybean 8 Day Old Root Infected With Cyst Nematode | src3c.pk004.c19 |
| wl1n | Wheat Leaf From 7 Day Old Seedling* | wl1n.pk0109.d5 wl1n.pk0110.c1 |
| wle1n | Wheat Leaf From 7 Day Old Etiolated Seedling* | wle1n.pk0075.b4 |
| wlm0 | Wheat Seedlings 0 Hour After Inoculation With <i>Erysiphe graminis f. sp tritici</i> | wlm0.pk0028.f4 |
| wlm96 | Wheat Seedlings 96 Hours After Inoculation With <i>Erysiphe graminis f. sp tritici</i> | wlm96.pk026.j1 wlm96.pk041.h3 wlm96.pk045.c11 |

*These libraries were normalized essentially as described in U.S. Pat. No. 5,482,845

**Application of 2-[(2,4-dihydro-2,6,9-trimethyl[1]benzothiopyrano[4,3-c]pyrazol-8-yl)carbonyl]-1,3-cyclohexanedione *S,S*-dioxide (synthesis and methods of using this compound are described in WO 97/19087, incorporated herein by reference) and 2-[(2,3-dihydro-5,8-dimethylspiro[4*H*-1-benzothiopyran-4,2'-[1,3]dioxolan]-6-yl)carbonyl]-1,3-cyclohexanedione *S,S*-dioxide (also named 2-[(2,3-dihydro-5,8-dimethylspiro[4*H*-1-benzothiopyran-4,2'-[1,3]dioxolan]-6-yl)carbonyl]-3-hydroxy-2-cyclohexen-1-one *S,S*-dioxide; synthesis and methods of using this compound are described in WO 97/01550, incorporated herein by reference).

10 ***Chemicals used included nitrilotriacetic acid, mercaptobenzothiazole, diethyldithiocarbamate

****For description of corn stages see Iowa State University Cooperative Extension Service Special Report No. 48.

15 cDNA libraries were prepared in Uni-ZAP™ XR vectors according to the manufacturer's protocol (Stratagene Cloning Systems, La Jolla, CA). Conversion of the Uni-ZAP™ XR libraries into plasmid libraries was accomplished according to the protocol provided by Stratagene. Upon conversion, cDNA inserts were contained in the plasmid vector pBluescript. cDNA inserts from randomly picked bacterial colonies containing
20 recombinant pBluescript plasmids were amplified via polymerase chain reaction using primers specific for vector sequences flanking the inserted cDNA sequences or plasmid DNA was prepared from cultured bacterial cells. Amplified insert DNAs or plasmid DNAs were sequenced in dye-primer sequencing reactions to generate partial cDNA sequences (expressed sequence tags or "ESTs"; see Adams, M. D. et al., (1991) *Science* 252:1651).
25 The resulting ESTs were analyzed using a Perkin Elmer Model 377 fluorescent sequencer.

EXAMPLE 2

Identification of cDNA Clones

ESTs encoding tryptophan biosynthetic enzymes were identified by conducting BLAST (Basic Local Alignment Search Tool; Altschul, S. F., et al., (1993) *J. Mol. Biol.*
30 215:403-410; see also www.ncbi.nlm.nih.gov/BLAST/) searches for similarity to sequences contained in the BLAST "nr" database (comprising all non-redundant GenBank CDS translations, sequences derived from the 3-dimensional structure Brookhaven Protein Data Bank, the last major release of the SWISS-PROT protein sequence database, EMBL, and DDBJ databases). The cDNA sequences obtained in Example 1 were analyzed for similarity
35 to all publicly available DNA sequences contained in the "nr" database using the BLASTN algorithm provided by the National Center for Biotechnology Information (NCBI). The DNA sequences were translated in all reading frames and compared for similarity to all publicly available protein sequences contained in the "nr" database using the BLASTX algorithm (Gish, W. and States, D. J. (1993) *Nature Genetics* 3:266-272) provided by the
40 NCBI. For convenience, the P-value (probability) of observing a match of a cDNA sequence to a sequence contained in the searched databases merely by chance as calculated by BLAST are reported herein as "pLog" values, which represent the negative of the

logarithm of the reported P-value. Accordingly, the greater the pLog value, the greater the likelihood that the cDNA sequence and the BLAST "hit" represent homologous proteins.

EXAMPLE 3

Characterization of cDNA Clones Encoding Anthranilate Synthase

5 The BLASTX search using the EST sequences from several corn, rice and wheat clones revealed similarity of the proteins encoded by the cDNAs to anthranilate synthase from *Arabidopsis thaliana* and *Ruta graveolens*. The BLASTX search using the nucleotide sequence from clone wre1n.pk0036.g10 also revealed similarity of the protein encoded by the cDNA to para-aminobenzoate synthase from *Streptomyces pristinaespiralis* (GenBank
10 Accession No.U60417; pLog = 27.39). In the process of comparing the corn ESTs it was found that clones cen3n.pk0210.c3, cta1n.pk0025.f8 and cen3n.pk0047.g5 had overlapping regions of homology. A comparison of the corn ESTs from clones cen3n.pk0212.h6 and cs1n.pk0017.d4 also had overlapping regions of homology. Using this homology it was possible to align the ESTs and assemble two contigs (a contig is an assemblage of
15 overlapping nucleic acid sequences to form one contiguous nucleotide sequence). The individual sequences were assembled into unique contiguous nucleotide sequences encoding anthranilate synthase from corn. The database accession numbers and BLAST results for each of these ESTs and contigs are shown in Table 3:

20

TABLE 3

BLAST Results for Clones Encoding Polypeptides Homologous to Anthranilate Synthase

| Clone | Organism | Database | |
|---|----------------------|----------------|------------------|
| | | Accession No | Blast Score pLog |
| Contig of: cen3n.pk0210.c3b cta1n.pk0025.f8 cen3n.pk0047.g5b | <i>A. thaliana</i> | GenBank L22585 | 70.69 |
| ceb3.pk0011.c12 | <i>R. graveolens</i> | GenBank L34344 | 48.15 |
| cta1n.pk0051.g3 | <i>R. graveolens</i> | GenBank L34344 | 32.22 |
| cbn2.pk0049.d2 | <i>R. graveolens</i> | GenBank L34344 | 35.00 |
| Contig of: cen3n.pk0212.h6 cs1n.pk0017.d4 | <i>A. thaliana</i> | GenBank L22585 | 34.00 |
| cco1n.pk0038.b1 | <i>A. thaliana</i> | GenBank L22585 | 17.69 |
| rls24.pk0002.e2 | <i>A. thaliana</i> | GenBank L22585 | 20.69 |
| wle1n.pk0075.b4 | <i>R. graveolens</i> | GenBank L34343 | 38.00 |

Characterization of cDNA Clones Encoding Anthranilate Synthase Alpha Subunit

25 The sequence of the entire cDNA insert in clone cta1n.pk0051.g3 was determined. A contig was assembled with this sequence and sequences from portions of the cDNA inserts

in clones cde1c.pk004.g2, p0125.czabf83r, p0005.cbmew77r, p0096.cnamt32r, p0032.crcas26r, p0031.ccmr07r, p0015.cdpfk12r and p0128.cpibl07r. The sequence of the entire cDNA insert in clone ctaln.pk0051.g3 was determined. A contig was assembled with this sequence and sequences from portions of the cDNA in clones p0115.clsmj70r, cbn2.pk0049.d2, cpl1c.pk008.m4 and p0102.ceraq01r. The BLASTX search using these contig sequences and the EST sequences from clones rcaln.pk004.p22, sdp3c.pk019.c6, sfl1.pk129.d10, wle1n.pk0075.b4, the 5' sequence from clone wlm96.pk026.j1 and the 3' sequence from clone wlm96.pk026.j1 revealed similarity of the protein encoded by the cDNA to anthranilate synthase alpha subunit from *Ruta graveolens* (NCBI General Identifier No. 960289). The BLAST results for each of these sequences are shown in Table 4:

TABLE 4

BLAST Results for Clones Encoding Polypeptides Homologous to Anthranilate Synthase Alpha Subunit

| Clone | BLAST pLog Score. NCBI General Identifier No. 960289 |
|--|---|
| Contig of: cde1c.pk004.g2 p0125.czabf83r p0005.cbmew77r p0096.cnamt32r p0032.crcas26r p0031.ccmr07r p0015.cdpfk12r p0128.cpibl07r ctaln.pk0051.g3 | >254 |
| Contig of: p0115.clsmj70r cbn2.pk0049.d2 cpl1c.pk008.m4 p0102.ceraq01r ceb3.pk0011.c12 | >254 |
| rcaln.pk004.p22 | 20.15 |
| sdp3c.pk019.c6 | 48.52 |
| sfl1.pk129.d10 | 34.70 |
| wle1n.pk0075.b4 (5'end) | 57.00 |
| wle1n.pk0075.b4 (3'end) | 35.70 |
| wlm96.pk026.j1 | 5.40 |

15

The sequence of the contig assembled from the entire cDNA insert from clone ctaln.pk0051.g3 and a portion of the cDNA insert in clones cde1c.pk004.g2, p0125.czabf83r, p0005.cbmew77r, p0096.cnamt32r, p0032.crcas26r, p0031.ccmr07r, p0015.cdpfk12r and p0128.cpibl07r is shown in SEQ ID NO:1; the deduced amino acid sequence of this cDNA is shown in SEQ ID NO:2. The amino acid sequence set forth in

20

SEQ ID NO:2 was evaluated by BLASTP, yielding a pLog value >254 versus the *Ruta graveolens* sequence. The sequence of the contig assembled from the entire cDNA insert in clone cta1n.pk0051.g3 and a portion of the cDNA in clones p0115.clsmj70r, cbn2.pk0049.d2, cpl1c.pk008.m4 and p0102.ceraq01r is shown in SEQ ID NO:3; the deduced amino acid sequence of this cDNA is shown in SEQ ID NO:4. The sequence of a portion of the cDNA insert from clone rca1n.pk004.p22 is shown in SEQ ID NO:5; the deduced amino acid sequence of this cDNA is shown in SEQ ID NO:6. The sequence of a portion of the cDNA insert from clone sdp3c.pk019.c6 is shown in SEQ ID NO:7; the deduced amino acid sequence of this cDNA is shown in SEQ ID NO:8. The sequence of a portion of the cDNA insert from clone sfl1.pk129.d10 is shown in SEQ ID NO:9; the deduced amino acid sequence of this cDNA is shown in SEQ ID NO:10. The sequence of the 5' terminal portion of the cDNA insert from clone wle1n.pk0075.b4 is shown in SEQ ID NO:11; the deduced amino acid sequence of this cDNA is shown in SEQ ID NO:12. The sequence of the 3' terminal portion of the cDNA insert from clone wle1n.pk0075.b4 is shown in SEQ ID NO:13; the deduced amino acid sequence of this cDNA is shown in SEQ ID NO:14. The sequence of a portion of the cDNA insert from clone wlm96.pk026.j1 is shown in SEQ ID NO:15; the deduced amino acid sequence of this cDNA is shown in SEQ ID NO:16.

Figure 2 presents an alignment of the amino acid sequence set forth in SEQ ID NO:2 and the *Ruta graveolens* anthranilate synthase alpha subunit sequence (SEQ ID NO:39). The amino acid sequence set forth in SEQ ID NO:2 is 64.7% similar to the *Ruta graveolens* sequence. Sequence alignments and percent similarity calculations were performed using the Megalign program of the LASARGENE bioinformatics computing suite (DNASTAR Inc., Madison, WI). Pairwise alignment of the sequences was performed using the Clustal method of alignment (Higgins, D.G. and Sharp, P.M. (1989) *CABIOS*. 5:151-153) with the default parameters (KTUPLE 1, GAP PENALTY=3, WINDOW=5 and DIAGONALS SAVED=5).

Sequence alignments and BLAST scores and probabilities indicate that the instant nucleic acid fragments encode an entire corn anthranilate synthase alpha subunit and portions of corn, rice, soybean and wheat anthranilate synthase alpha subunits. These sequences represent the first corn, rice, soybean and wheat sequences encoding anthranilate synthase alpha subunit.

Characterization of cDNA Clones Encoding Anthranilate Synthase Beta Subunit

The sequence of a larger portion of the cDNA insert in clone cen3n.pk0210.c3 was determined, this sequence includes the sequence from clone cta1n.pk0025.f8. A contig was assembled with the sequence from clone cen3n.pk0210.c3 and sequence from a portion of the cDNA in clones cepe7.pk0012.g2 and cen3n.pk0047.g5. The BLASTX search using these contig sequences, the sequences from the contig assembled of clones p0126.cnlcu75r, cen3n.pk0212.h6, cco1n.pk0038.b1, csi1n.pk0017.d4, p0031.ccmaj08r and ctn1c.pk001.110,

the contig assembled from clones rds2c.pk004.i9, rls24.pk0002.e2 and rl0n.pk113.b4 and the EST sequences from clone wlm96.pk045.c11 revealed similarity of the proteins encoded by the cDNAs to anthranilate synthase beta subunit from *Arabidopsis thaliana* (NCBI General Identifier No. 541849). The BLAST results for each of these ESTs are shown in Table 5:

5

TABLE 5

BLAST Results for Clones Encoding Polypeptides Homologous
to Anthranilate Synthase Beta Subunit

| Clone | BLAST pLog Score NCBI General Identifier No. 541849 |
|--|--|
| Contig of: cen3n.pk0210.c3 cepe7.pk0012.g2 cen3n.pk0047.g5 | 84.52 |
| Contig of: p0126.cnlcu75r cen3n.pk0212.h6 cco1n.pk0038.b1 csi1n.pk0017.d4 p0031.ccmαι08r ctn1c.pk001.110 | 110.0 |
| Contig of: rds2c.pk004.i9 rls24.pk0002.e2 rl0n.pk113.b4 | 20.15 |
| wlm96.pk045.c11 | 15.00 |

- 10 The sequence of the contig assembled from a portion of the cDNA insert from clones cen3n.pk0210.c3, cepe7.pk0012.g2 and cen3n.pk0047.g5 is shown in SEQ ID NO:17; the deduced amino acid sequence of this cDNA is shown in SEQ ID NO:18. The sequence of the contig assembled from a portion of the cDNA insert from clones p0126.cnlcu75r, cen3n.pk0212.h6, cco1n.pk0038.b1, csi1n.pk0017.d4, p0031.ccmαι08r and ctn1c.pk001.110
- 15 is shown in SEQ ID NO:17; the deduced amino acid sequence of this cDNA is shown in SEQ ID NO:18. The amino acid sequence set forth in SEQ ID NO:18 was evaluated by BLASTP, yielding a pLog value of 92.22 versus the *Arabidopsis thaliana* sequence. The sequence of the contig assembled from a portion of the cDNA insert from clones rds2c.pk004.i9, rls24.pk0002.e2 and rl0n.pk113.b4 is shown in SEQ ID NO:19; the deduced
- 20 amino acid sequence of this cDNA is shown in SEQ ID NO:20. The sequence of a portion of the cDNA insert from clone wlm96.pk045.c11 is shown in SEQ ID NO:21; the deduced amino acid sequence of this cDNA is shown in SEQ ID NO:22.

Figure 3 presents an alignment of the amino acid sequences set forth in SEQ ID NOs:20 and 22 and the *Arabidopsis thaliana* anthranilate synthase beta subunit sequence (SEQ ID NO:40). The amino acid sequences set forth in SEQ ID NO:20 are 58.0% similar

25 to the *Arabidopsis thaliana* sequence while the amino acid sequences set forth in SEQ ID

NO:22 are 57.1% similar to the *Arabidopsis thaliana* sequence. Sequence alignments and percent similarity calculations were performed using the Megalign program of the LASARGENE bioinformatics computing suite (DNASTAR Inc., Madison, WI). Multiple alignment of the sequences was performed using the Clustal method of alignment (Higgins, D.G. and Sharp, P.M. (1989) *CABIOS*. 5:151-153) with the default parameters (GAP PENALTY=10, GAP LENGTH PENALTY=10).

Sequence alignments and BLAST scores and probabilities indicate that the instant nucleic acid fragments encode an entire corn anthranilate synthase beta subunit, an almost entire rice anthranilate synthase beta subunit and portions of corn and wheat anthranilate synthase beta subunit. These sequences represent the first corn, rice and wheat sequences encoding anthranilate synthase beta subunit.

EXAMPLE 4

Characterization of cDNA Clones Encoding Tryptophan Synthase

The BLASTX search using the EST sequences from several corn, rice, soybean and wheat clones revealed similarity of the proteins encoded by the cDNAs to tryptophan synthase from *Zea mays* and *Arabidopsis thaliana*. In the process of comparing the rice ESTs it was found that clones wl1n.pk0109.d5 and wl1n.pk0110.c1 had overlapping regions of homology. A comparison of the corn ESTs from clones chp2.pk0020.e2, cen3n.pk0147.d4, lkr.pk0013.g1, cs1.pk0089.a8, ctal1n.pk0057.d6, cen3n.pk0014.c5, cen3n.pk0193.c5 and cen3n.pk0135.d1 also revealed overlapping regions of homology. A comparison of the corn ESTs from clones m.15.6.g07.sk20, m.15.4.g07.sk20 and cco1n.pk0030.g7 revealed overlapping regions of homology. Using this homology it was possible to align the ESTs and assemble four contigs (a contig is an assemblage of overlapping nucleic acid sequences to form one contiguous nucleotide sequence). The individual sequences were assembled into unique contiguous nucleotide sequences encoding tryptophan synthase from corn, rice, soybean and wheat. The database accession numbers and BLAST results for each of these ESTs and contigs are shown in Table 6:

TABLE 6

BLASTX Results for Clones Encoding Polypeptides Homologous to Tryptophan Synthase

| Clone | Organism | Database Accession No | Blast Score pLog |
|--|--------------------|-----------------------|------------------|
| rr1.pk0038.h6 | <i>A. thaliana</i> | GenBank U18993 | 26.00 |
| Contig of: wl1n.pk0109.d5 wl1n.pk0110.c1 | <i>Z. mays</i> | GenBank X76713 | 52.39 |
| wl1n.pk0067.f2 | <i>A. thaliana</i> | GenBank U18993 | 19.69 |
| cr1n.pk0033.f8 | <i>Z. mays</i> | GenBank X76713 | 47.69 |
| cr1n.pk0052.b8 | <i>Z. mays</i> | GenBank X76713 | 25.70 |

| Clone | Organism | Database Accession No | Blast Score pLog |
|---|----------------|--------------------------|------------------|
| Contig of: chp2.pk0020.e2 cen3n.pk0147.d4 lkr.pk0013.g1 cs1.pk0089.a8 cta1n.pk0057.d6 cen3n.pk0014.c5 cen3n.pk0193.c5 cen3n.pk0135.d1 | <i>Z. mays</i> | GenBank X76713 | 83.40 |
| Contig of: m.15.6.g07.sk20 m15.4.g07.sk20 cco1n.pk0030.g7 | <i>Z. mays</i> | GenBank X76713 | 25.52 |

Characterization of cDNA Clones Encoding Tryptophan Synthase Alpha Subunit

The sequence of the entire cDNA insert in clone lkr.pk0013.g1 and a larger portion of the sequence from clone chp2.pk0020.e2 have been determined. A contig has been assembled with these two sequences and they have been found to include the sequences from clones cco1n.pk0030.g7, m.15.6.g07.sk20, cen3n.pk0014.c5, cen3n.pk0135.d1, cen3n.pk0147.d4, cen3n.pk0193.c5, cta1n.pk0057.d6, cs1.pk0089.a8 and m.15.6.g07.sk20. The sequence from the entire cDNA insert in clone cr1n.pk0052.b8 has been determined, it includes the sequences from clone cr1n.pk0033.f8. The BLASTX search using these sequences, the sequence from the contig assembled of clones res1c.pk004.c4 and rr1.pk0071.f2, the EST sequences from clone rr1.pk0038.h6 and the sequence of the contig assembled from clones src3c.pk004.c19 and sls2c.pk025.d11 revealed similarity of the proteins encoded by the cDNAs to tryptophan synthase alpha subunit from *Arabidopsis thaliana* (NCBI General Identifier No. 2129755). The BLASTX search using the sequence of the contig assembled from clones wl1n.pk0109.d5 and wl1n.pk0110.c1 revealed similarity of the proteins encoded by the cDNAs to tryptophan synthase alpha subunit from *Zea mays* (NCBI General Identifier No. 1174783). The BLAST results for each of these sequences are shown in Table 7:

TABLE 7

BLAST Results for Clones Encoding Polypeptides Homologous
to Tryptophan Synthase Alpha Subunit

| Clone | NCBI General Identifier No. | BLAST pLog Score |
|--|-----------------------------|------------------|
| Contig of: chp2.pk0020.e2 lkr.pk0013.g1 | 2129755 | 124.0 |
| cr1n.pk0052.b8 | 2129755 | 120.0 |
| Contig of: res1c.pk004.c4 rr1.pk0071.f2 | 2129755 | 78.70 |
| rr1.pk0038.h6 | 2129755 | 125.0 |
| Contig of: src3c.pk004.c19 sls2c.pk025.d11 | 2129755 | 49.00 |
| Contig of: wlm0.pk0028.f4 wlm96.pk041.h3 | 1174783 | 33.70 |
| Contig of: wl1n.pk0109.d5 wl1n.pk0110.c1 | 1174783 | 85.70 |

- 5 The sequence of the contig assembled from the entire cDNA insert in clone lkr.pk0013.g1 and a large portion of the cDNA insert in clone chp2.pk0020.e2 is shown in SEQ ID NO:25; the deduced amino acid sequence of this cDNA is shown in SEQ ID NO:26. The amino acid sequence set forth in SEQ ID NO:26 was evaluated by BLASTP, yielding a pLog value of 105.0 versus the *Arabidopsis thaliana* sequence. The sequence of the entire
- 10 cDNA insert in clone cr1n.pk0052.b8 was determined and is shown in SEQ ID NO:27; the deduced amino acid sequence of this cDNA is shown in SEQ ID NO:28. The amino acid sequence set forth in SEQ ID NO:28 was evaluated by BLASTP, yielding a pLog value of 98.22 versus the *Arabidopsis thaliana* sequence. The sequence of the contig assembled from a portion of the cDNA insert in clones res1c.pk004.c4 and rr1.pk0071.f2 is shown in SEQ
- 15 ID NO:29; the deduced amino acid sequence of this cDNA is shown in SEQ ID NO:30. The sequence of the entire cDNA insert in clone rr1.pk0038.h6 was determined and is shown in SEQ ID NO:31; the deduced amino acid sequence of this cDNA is shown in SEQ ID NO:32. The amino acid sequence set forth in SEQ ID NO:32 was evaluated by BLASTP, yielding a pLog value of 106.0 versus the *Arabidopsis thaliana* sequence. The sequence of the contig
- 20 assembled from a portion of the cDNA insert from clones src3c.pk004.c19 and sls2c.pk025.d11 is shown in SEQ ID NO:33; the deduced amino acid sequence of this cDNA is shown in SEQ ID NO:34. The sequence of the contig assembled from a portion of the cDNA insert from clones wlm0.pk0028.f4 and wlm96.pk041.h3 is shown in SEQ ID NO:35; the deduced amino acid sequence of this cDNA is shown in SEQ ID NO:36. The

sequence of the contig assembled from a portion of the cDNA insert from clones wl1n.pk0109.d5 and wl1n.pk0110.c1 is shown in SEQ ID NO:37; the deduced amino acid sequence of this cDNA is shown in SEQ ID NO:38.

Figure 3 presents an alignment of the amino acid sequences set forth in SEQ ID NOs:26, 28 and 32 and the *Arabidopsis thaliana* sequence (SEQ ID NO:41). The data in Table 8 represents a calculation of the percent identity of the amino acid sequences set forth in SEQ ID NOs:26, 28 and 32 and the *Arabidopsis thaliana* tryptophan synthase alpha subunit sequence.

TABLE 8
Percent Identity of Amino Acid Sequences Deduced From the Nucleotide Sequences of cDNA Clones Encoding Polypeptides Homologous to Tryptophan Synthase Alpha Subunit

| Clone | SEQ ID NO. | Percent Identity to 2129755 |
|---|------------|-----------------------------|
| Contig of: chp2.pk0020.e2 lkr.pk0013.g1 | 26 | 59.0 |
| cr1n.pk0052.b8 | 28 | 56.7 |
| rr1.pk0038.h6 | 32 | 59.9 |

Sequence alignments and percent similarity calculations were performed using the Megalign program of the LASARGENE bioinformatics computing suite (DNASTAR Inc., Madison, WI). Multiple alignment of the sequences was performed using the Clustal method of alignment (Higgins, D.G. and Sharp, P.M. (1989) *CABIOS*. 5:151-153) with the default parameters (GAP PENALTY=10, GAP LENGTH PENALTY=10).

Sequence alignments and BLAST scores and probabilities indicate that the instant nucleic acid fragments encode entire or nearly entire corn and rice tryptophan synthase alpha subunit and portions of rice, soybean and wheat tryptophan synthase alpha subunit. These sequences represent the first rice, soybean and wheat and variant corn sequences encoding tryptophan synthase alpha subunit.

EXAMPLE 5 Expression of Chimeric Genes in Monocot Cells

A chimeric gene comprising a cDNA encoding a tryptophan biosynthetic enzyme in sense orientation with respect to the maize 27 kD zein promoter that is located 5' to the cDNA fragment, and the 10 kD zein 3' end that is located 3' to the cDNA fragment, can be constructed. The cDNA fragment of this gene may be generated by polymerase chain reaction (PCR) of the cDNA clone using appropriate oligonucleotide primers. Cloning sites (NcoI or SmaI) can be incorporated into the oligonucleotides to provide proper orientation of the DNA fragment when inserted into the digested vector pML103 as described below. Amplification is then performed in a standard PCR. The amplified DNA is then digested

with restriction enzymes NcoI and SmaI and fractionated on an agarose gel. The appropriate band can be isolated from the gel and combined with a 4.9 kb NcoI-SmaI fragment of the plasmid pML103. Plasmid pML103 has been deposited under the terms of the Budapest Treaty at ATCC (American Type Culture Collection, 10801 University Blvd., Manassas, VA
5 20110-2209), and bears accession number ATCC 97366. The DNA segment from pML103 contains a 1.05 kb SalI-NcoI promoter fragment of the maize 27 kD zein gene and a 0.96 kb SmaI-SalI fragment from the 3' end of the maize 10 kD zein gene in the vector pGem9Zf(+) (Promega). Vector and insert DNA can be ligated at 15°C overnight, essentially as described (Maniatis). The ligated DNA may then be used to transform *E. coli* XL1-Blue
10 (Epicurian Coli XL-1 Blue™; Stratagene). Bacterial transformants can be screened by restriction enzyme digestion of plasmid DNA and limited nucleotide sequence analysis using the dideoxy chain termination method (Sequenase™ DNA Sequencing Kit; U. S. Biochemical). The resulting plasmid construct would comprise a chimeric gene encoding, in the 5' to 3' direction, the maize 27 kD zein promoter, a cDNA fragment encoding a
15 tryptophan biosynthetic enzyme, and the 10 kD zein 3' region.

The chimeric gene described above can then be introduced into corn cells by the following procedure. Immature corn embryos can be dissected from developing caryopses derived from crosses of the inbred corn lines H99 and LH132. The embryos are isolated 10 to 11 days after pollination when they are 1.0 to 1.5 mm long. The embryos are then placed
20 with the axis-side facing down and in contact with agarose-solidified N6 medium (Chu et al., (1975) *Sci. Sin. Peking* 18:659-668). The embryos are kept in the dark at 27°C. Friable embryogenic callus consisting of undifferentiated masses of cells with somatic proembryoids and embryoids borne on suspensor structures proliferates from the scutellum of these immature embryos. The embryogenic callus isolated from the primary explant can
25 be cultured on N6 medium and sub-cultured on this medium every 2 to 3 weeks.

The plasmid, p35S/Ac (obtained from Dr. Peter Eckes, Hoechst Ag, Frankfurt, Germany) may be used in transformation experiments in order to provide for a selectable marker. This plasmid contains the *Pat* gene (see European Patent Publication 0 242 236) which encodes phosphinothricin acetyl transferase (PAT). The enzyme PAT confers
30 resistance to herbicidal glutamine synthetase inhibitors such as phosphinothricin. The *pat* gene in p35S/Ac is under the control of the 35S promoter from Cauliflower Mosaic Virus (Odell et al. (1985) *Nature* 313:810-812) and the 3' region of the nopaline synthase gene from the T-DNA of the Ti plasmid of *Agrobacterium tumefaciens*.

The particle bombardment method (Klein et al., (1987) *Nature* 327:70-73) may be
35 used to transfer genes to the callus culture cells. According to this method, gold particles (1 µm in diameter) are coated with DNA using the following technique. Ten µg of plasmid DNAs are added to 50 µL of a suspension of gold particles (60 mg per mL). Calcium chloride (50 µL of a 2.5 M solution) and spermidine free base (20 µL of a 1.0 M solution) are added to the particles. The suspension is vortexed during the addition of these solutions.

After 10 minutes, the tubes are briefly centrifuged (5 sec at 15,000 rpm) and the supernatant removed. The particles are resuspended in 200 μ L of absolute ethanol, centrifuged again and the supernatant removed. The ethanol rinse is performed again and the particles resuspended in a final volume of 30 μ L of ethanol. An aliquot (5 μ L) of the DNA-coated
5 gold particles can be placed in the center of a Kapton™ flying disc (Bio-Rad Labs). The particles are then accelerated into the corn tissue with a Biolistic™ PDS-1000/He (Bio-Rad Instruments, Hercules CA), using a helium pressure of 1000 psi, a gap distance of 0.5 cm and a flying distance of 1.0 cm.

For bombardment, the embryogenic tissue is placed on filter paper over agarose-
10 solidified N6 medium. The tissue is arranged as a thin lawn and covered a circular area of about 5 cm in diameter. The petri dish containing the tissue can be placed in the chamber of the PDS-1000/He approximately 8 cm from the stopping screen. The air in the chamber is then evacuated to a vacuum of 28 inches of Hg. The macrocarrier is accelerated with a helium shock wave using a rupture membrane that bursts when the He pressure in the shock
15 tube reaches 1000 psi.

Seven days after bombardment the tissue can be transferred to N6 medium that contains gluphosinate (2 mg per liter) and lacks casein or proline. The tissue continues to grow slowly on this medium. After an additional 2 weeks the tissue can be transferred to
20 fresh N6 medium containing gluphosinate. After 6 weeks, areas of about 1 cm in diameter of actively growing callus can be identified on some of the plates containing the glufosinate-supplemented medium. These calli may continue to grow when sub-cultured on the selective medium.

Plants can be regenerated from the transgenic callus by first transferring clusters of tissue to N6 medium supplemented with 0.2 mg per liter of 2,4-D. After two weeks the
25 tissue can be transferred to regeneration medium (Fromm et al., (1990) *Bio/Technology* 8:833-839).

EXAMPLE 6

Expression of Chimeric Genes in Dicot Cells

A seed-specific expression cassette composed of the promoter and transcription
30 terminator from the gene encoding the β subunit of the seed storage protein phaseolin from the bean *Phaseolus vulgaris* (Doyle et al. (1986) *J. Biol. Chem.* 261:9228-9238) can be used for expression of the instant tryptophan biosynthetic enzyme in transformed soybean. The phaseolin cassette includes about 500 nucleotides upstream (5') from the translation initiation codon and about 1650 nucleotides downstream (3') from the translation stop codon of
35 phaseolin. Between the 5' and 3' regions are the unique restriction endonuclease sites Nco I (which includes the ATG translation initiation codon), Sma I, Kpn I and Xba I. The entire cassette is flanked by Hind III sites.

The cDNA fragment of this gene may be generated by polymerase chain reaction (PCR) of the cDNA clone using appropriate oligonucleotide primers. Cloning sites can be

incorporated into the oligonucleotides to provide proper orientation of the DNA fragment when inserted into the expression vector. Amplification is then performed as described above, and the isolated fragment is inserted into a pUC18 vector carrying the seed expression cassette.

- 5 Soybean embryos may then be transformed with the expression vector comprising sequences encoding a tryptophan biosynthetic enzyme. To induce somatic embryos, cotyledons, 3-5 mm in length dissected from surface sterilized, immature seeds of the soybean cultivar A2872, can be cultured in the light or dark at 26°C on an appropriate agar medium for 6-10 weeks. Somatic embryos which produce secondary embryos are then
10 excised and placed into a suitable liquid medium. After repeated selection for clusters of somatic embryos which multiplied as early, globular staged embryos, the suspensions are maintained as described below.

- Soybean embryogenic suspension cultures can maintained in 35 mL liquid media on a rotary shaker, 150 rpm, at 26°C with florescent lights on a 16:8 hour day/night schedule.
15 Cultures are subcultured every two weeks by inoculating approximately 35 mg of tissue into 35 mL of liquid medium.

- Soybean embryogenic suspension cultures may then be transformed by the method of particle gun bombardment (Kline et al. (1987) *Nature* (London) 327:70, U.S. Patent No. 4,945,050). A DuPont Biolistic™ PDS1000/HE instrument (helium retrofit) can be used
20 for these transformations.

- A selectable marker gene which can be used to facilitate soybean transformation is a chimeric gene composed of the 35S promoter from Cauliflower Mosaic Virus (Odell et al.(1985) *Nature* 313:810-812), the hygromycin phosphotransferase gene from plasmid pJR225 (from *E. coli*; Gritz et al.(1983) *Gene* 25:179-188) and the 3' region of the nopaline synthase gene from the T-DNA of the Ti plasmid of *Agrobacterium tumefaciens*. The seed
25 expression cassette comprising the phaseolin 5' region, the fragment encoding the tryptophan biosynthetic enzyme and the phaseolin 3' region can be isolated as a restriction fragment. This fragment can then be inserted into a unique restriction site of the vector carrying the marker gene.

- 30 To 50 µL of a 60 mg/mL 1 µm gold particle suspension is added (in order): 5 µL DNA (1 µg/µL), 20 µl spermidine (0.1 M), and 50 µL CaCl₂ (2.5 M). The particle preparation is then agitated for three minutes, spun in a microfuge for 10 seconds and the supernatant removed. The DNA-coated particles are then washed once in 400 µL 70% ethanol and resuspended in 40 µL of anhydrous ethanol. The DNA/particle suspension can
35 be sonicated three times for one second each. Five µL of the DNA-coated gold particles are then loaded on each macro carrier disk.

Approximately 300-400 mg of a two-week-old suspension culture is placed in an empty 60x15 mm petri dish and the residual liquid removed from the tissue with a pipette. For each transformation experiment, approximately 5-10 plates of tissue are normally

bombarded. Membrane rupture pressure is set at 1100 psi and the chamber is evacuated to a vacuum of 28 inches mercury. The tissue is placed approximately 3.5 inches away from the retaining screen and bombarded three times. Following bombardment, the tissue can be divided in half and placed back into liquid and cultured as described above.

- 5 Five to seven days post bombardment, the liquid media may be exchanged with fresh media, and eleven to twelve days post bombardment with fresh media containing 50 mg/mL hygromycin. This selective media can be refreshed weekly. Seven to eight weeks post bombardment, green, transformed tissue may be observed growing from untransformed, necrotic embryogenic clusters. Isolated green tissue is removed and inoculated into individual flasks to generate new, clonally propagated, transformed embryogenic suspension cultures. Each new line may be treated as an independent transformation event. These suspensions can then be subcultured and maintained as clusters of immature embryos or regenerated into whole plants by maturation and germination of individual somatic embryos.

EXAMPLE 7

15 Expression of Chimeric Genes in Microbial Cells

- The cDNAs encoding the instant tryptophan biosynthetic enzymes can be inserted into the T7 *E. coli* expression vector pBT430. This vector is a derivative of pET-3a (Rosenberg et al. (1987) *Gene* 56:125-135) which employs the bacteriophage T7 RNA polymerase/T7 promoter system. Plasmid pBT430 was constructed by first destroying the EcoR I and Hind III sites in pET-3a at their original positions. An oligonucleotide adaptor containing EcoR I and Hind III sites was inserted at the BamH I site of pET-3a. This created pET-3aM with additional unique cloning sites for insertion of genes into the expression vector. Then, the Nde I site at the position of translation initiation was converted to an Nco I site using oligonucleotide-directed mutagenesis. The DNA sequence of pET-3aM in this region, 25 5'-CATATGG, was converted to 5'-CCCATGG in pBT430.

- Plasmid DNA containing a cDNA may be appropriately digested to release a nucleic acid fragment encoding the protein. This fragment may then be purified on a 1% NuSieve GTG™ low melting agarose gel (FMC). Buffer and agarose contain 10 µg/ml ethidium bromide for visualization of the DNA fragment. The fragment can then be purified from the agarose gel by digestion with GELase™ (Epicentre Technologies) according to the manufacturer's instructions, ethanol precipitated, dried and resuspended in 20 µL of water. Appropriate oligonucleotide adapters may be ligated to the fragment using T4 DNA ligase (New England Biolabs, Beverly, MA). The fragment containing the ligated adapters can be purified from the excess adapters using low melting agarose as described above. The vector 35 pBT430 is digested, dephosphorylated with alkaline phosphatase (NEB) and deproteinized with phenol/chloroform as described above. The prepared vector pBT430 and fragment can then be ligated at 16°C for 15 hours followed by transformation into DH5 electrocompetent cells (GIBCO BRL). Transformants can be selected on agar plates containing LB media and 100 µg/mL ampicillin. Transformants containing the gene encoding the tryptophan

biosynthetic enzyme are then screened for the correct orientation with respect to the T7 promoter by restriction enzyme analysis.

For high level expression, a plasmid clone with the cDNA insert in the correct orientation relative to the T7 promoter can be transformed into *E. coli* strain BL21(DE3) (Studier et al. (1986) *J. Mol. Biol.* 189:113-130). Cultures are grown in LB medium containing ampicillin (100 mg/L) at 25°C. At an optical density at 600 nm of approximately 1, IPTG (isopropylthio- β -galactoside, the inducer) can be added to a final concentration of 0.4 mM and incubation can be continued for 3 h at 25°. Cells are then harvested by centrifugation and re-suspended in 50 μ L of 50 mM Tris-HCl at pH 8.0 containing 0.1 mM DTT and 0.2 mM phenyl methylsulfonyl fluoride. A small amount of 1 mm glass beads can be added and the mixture sonicated 3 times for about 5 seconds each time with a microprobe sonicator. The mixture is centrifuged and the protein concentration of the supernatant determined. One μ g of protein from the soluble fraction of the culture can be separated by SDS-polyacrylamide gel electrophoresis. Gels can be observed for protein bands migrating at the expected molecular weight.

EXAMPLE 8

Evaluating Compounds for Their Ability to Inhibit the Activity of Tryptophan Biosynthetic Enzymes

The tryptophan biosynthetic enzymes described herein may be produced using any number of methods known to those skilled in the art. Such methods include, but are not limited to, expression in bacteria as described in Example 7, or expression in eukaryotic cell culture, *in planta*, and using viral expression systems in suitably infected organisms or cell lines. The instant tryptophan biosynthetic enzymes may be expressed either as mature forms of the proteins as observed *in vivo* or as fusion proteins by covalent attachment to a variety of enzymes, proteins or affinity tags. Common fusion protein partners include glutathione S-transferase ("GST"), thioredoxin ("Trx"), maltose binding protein, and C- and/or N-terminal hexahistidine polypeptide ("His)₆"). The fusion proteins may be engineered with a protease recognition site at the fusion point so that fusion partners can be separated by protease digestion to yield intact mature enzyme. Examples of such proteases include thrombin, enterokinase and factor Xa. However, any protease can be used which specifically cleaves the peptide connecting the fusion protein and the enzyme.

Purification of the instant tryptophan biosynthetic enzymes, if desired, may utilize any number of separation technologies familiar to those skilled in the art of protein purification. Examples of such methods include, but are not limited to, homogenization, filtration, centrifugation, heat denaturation, ammonium sulfate precipitation, desalting, pH precipitation, ion exchange chromatography, hydrophobic interaction chromatography and affinity chromatography, wherein the affinity ligand represents a substrate, substrate analog or inhibitor. When the tryptophan biosynthetic enzymes are expressed as fusion proteins, the purification protocol may include the use of an affinity resin which is specific for the fusion

protein tag attached to the expressed enzyme or an affinity resin containing ligands which are specific for the enzyme. For example, a tryptophan biosynthetic enzyme may be expressed as a fusion protein coupled to the C-terminus of thioredoxin. In addition, a (His)₆ peptide may be engineered into the N-terminus of the fused thioredoxin moiety to afford additional opportunities for affinity purification. Other suitable affinity resins could be synthesized by linking the appropriate ligands to any suitable resin such as Sepharose-4B. In an alternate embodiment, a thioredoxin fusion protein may be eluted using dithiothreitol; however, elution may be accomplished using other reagents which interact to displace the thioredoxin from the resin. These reagents include β -mercaptoethanol or other reduced thiol. The eluted fusion protein may be subjected to further purification by traditional means as stated above, if desired. Proteolytic cleavage of the thioredoxin fusion protein and the enzyme may be accomplished after the fusion protein is purified or while the protein is still bound to the ThioBond™ affinity resin or other resin.

Crude, partially purified or purified enzyme, either alone or as a fusion protein, may be utilized in assays for the evaluation of compounds for their ability to inhibit enzymatic activation of the tryptophan biosynthetic enzymes disclosed herein. Assays may be conducted under well known experimental conditions which permit optimal enzymatic activity. For example, assays for anthranilate synthase alpha subunit are presented by Bohlmann J et al. (1995) *Plant J* 7:491-501. Assays for anthranilate synthase beta subunit are presented by Walker M. S. and DeMoss J. A. (1983) *J Biol Chem* 258:3571-3575. Assays for tryptophan synthase alpha subunit are presented by Zhao J and Last R. L. (1995) *J Biol Chem* 270:6081-7.

CLAIMS

What is claimed is:

1. An isolated nucleic acid fragment encoding all or a substantial portion of an anthranilate synthase alpha subunit comprising a member selected from the group consisting of:
 - (a) an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in a member selected from the group consisting of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14 and SEQ ID NO:16;
 - (b) an isolated nucleic acid fragment that is substantially similar to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in a member selected from the group consisting of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14 and SEQ ID NO:16; and
 - (c) an isolated nucleic acid fragment that is complementary to (a) or (b).
2. The isolated nucleic acid fragment of Claim 1 wherein the isolated nucleic acid fragment hybridizes under stringent conditions to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:2.
3. The isolated nucleic acid fragment of Claim 1 wherein the isolated nucleic acid fragment hybridizes under stringent conditions to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:4.
4. The isolated nucleic acid fragment of Claim 1 wherein the isolated nucleic acid fragment hybridizes under stringent conditions to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:6.
5. The isolated nucleic acid fragment of Claim 1 wherein the isolated nucleic acid fragment hybridizes under stringent conditions to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:8.
6. The isolated nucleic acid fragment of Claim 1 wherein the isolated nucleic acid fragment hybridizes under stringent conditions to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:10.
7. The isolated nucleic acid fragment of Claim 1 wherein the isolated nucleic acid fragment hybridizes under stringent conditions to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:12.
8. The isolated nucleic acid fragment of Claim 1 wherein the isolated nucleic acid fragment hybridizes under stringent conditions to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:14.

9. The isolated nucleic acid fragment of Claim 1 wherein the isolated nucleic acid fragment hybridizes under stringent conditions to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:16.
10. The isolated nucleic acid fragment of Claim 1 wherein the isolated nucleic acid
5 fragment is at least 80% similar to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:2.
11. The isolated nucleic acid fragment of Claim 1 wherein the isolated nucleic acid fragment is at least 80% similar to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:4.
- 10 12. The isolated nucleic acid fragment of Claim 1 wherein the isolated nucleic acid fragment is at least 80% similar to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:6.
13. The isolated nucleic acid fragment of Claim 1 wherein the isolated nucleic acid fragment is at least 80% similar to an isolated nucleic acid fragment encoding all or a
15 substantial portion of the amino acid sequence set forth in SEQ ID NO:8.
14. The isolated nucleic acid fragment of Claim 1 wherein the isolated nucleic acid fragment is at least 80% similar to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:10.
15. The isolated nucleic acid fragment of Claim 1 wherein the isolated nucleic acid
20 fragment is at least 80% similar to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:12.
16. The isolated nucleic acid fragment of Claim 1 wherein the isolated nucleic acid fragment is at least 80% similar to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:14.
- 25 17. The isolated nucleic acid fragment of Claim 1 wherein the isolated nucleic acid fragment is at least 80% similar to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:16.
18. The isolated nucleic acid fragment of Claim 1 wherein the nucleotide sequence of the fragment comprises all or a portion of the sequence set forth in a member selected
30 from the group consisting of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13 and SEQ ID NO:15.
19. A chimeric gene comprising the nucleic acid fragment of Claim 1 operably linked to suitable regulatory sequences.
20. A transformed host cell comprising the chimeric gene of Claim 19.
- 35 21. An anthranilate synthase alpha subunit polypeptide comprising all or a substantial portion of the amino acid sequence set forth in a member selected from the group consisting of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14 and SEQ ID NO:16.

22. An isolated nucleic acid fragment encoding all or a substantial portion of an anthranilate synthase beta subunit comprising a member selected from the group consisting of:

- 5 (a) an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in a member selected from the group consisting of SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22 and SEQ ID NO:24;
- 10 (b) an isolated nucleic acid fragment that is substantially similar to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in a member selected from the group consisting of SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22 and SEQ ID NO:24; and
- (c) an isolated nucleic acid fragment that is complementary to (a) or (b).

23. The isolated nucleic acid fragment of Claim 22 wherein the isolated nucleic acid fragment hybridizes under stringent conditions to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:18.

24. The isolated nucleic acid fragment of Claim 22 wherein the isolated nucleic acid fragment hybridizes under stringent conditions to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:20.

20 25. The isolated nucleic acid fragment of Claim 22 wherein the isolated nucleic acid fragment hybridizes under stringent conditions to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:22.

25 26. The isolated nucleic acid fragment of Claim 22 wherein the isolated nucleic acid fragment hybridizes under stringent conditions to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:24.

27. The isolated nucleic acid fragment of Claim 22 wherein the isolated nucleic acid fragment is at least 80% similar to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:18.

30 28. The isolated nucleic acid fragment of Claim 22 wherein the isolated nucleic acid fragment is at least 80% similar to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:20.

29. The isolated nucleic acid fragment of Claim 22 wherein the isolated nucleic acid fragment is at least 80% similar to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:22.

35 30. The isolated nucleic acid fragment of Claim 22 wherein the isolated nucleic acid fragment is at least 80% similar to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:24.

31. The isolated nucleic acid fragment of Claim 22 wherein the nucleotide sequence of the fragment comprises all or a portion of the sequence set forth in a member

selected from the group consisting of SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21 and SEQ ID NO:23.

32. A chimeric gene comprising the nucleic acid fragment of Claim 22 operably linked to suitable regulatory sequences.

5 33. A transformed host cell comprising the chimeric gene of Claim 32.

34. An anthranilate synthase beta subunit polypeptide comprising all or a substantial portion of the amino acid sequence set forth in a member selected from the group consisting of SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22 and SEQ ID NO:24.

10 35. An isolated nucleic acid fragment encoding all or a substantial portion of a tryptophan synthase alpha subunit comprising a member selected from the group consisting of:

(a) an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in a member selected from the group consisting of SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36 and SEQ ID NO:38;

(b) an isolated nucleic acid fragment that is substantially similar to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in a member selected from the group consisting of SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36 and SEQ ID NO:38; and

(c) an isolated nucleic acid fragment that is complementary to (a) or (b).

36. The isolated nucleic acid fragment of Claim 35 wherein the isolated nucleic acid fragment hybridizes under stringent conditions to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:26.

25 37. The isolated nucleic acid fragment of Claim 35 wherein the isolated nucleic acid fragment hybridizes under stringent conditions to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:28.

38. The isolated nucleic acid fragment of Claim 35 wherein the isolated nucleic acid fragment hybridizes under stringent conditions to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:30.

39. The isolated nucleic acid fragment of Claim 35 wherein the isolated nucleic acid fragment hybridizes under stringent conditions to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:32.

40. The isolated nucleic acid fragment of Claim 35 wherein the isolated nucleic acid fragment hybridizes under stringent conditions to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:34.

41. The isolated nucleic acid fragment of Claim 35 wherein the isolated nucleic acid fragment hybridizes under stringent conditions to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:36.

42. The isolated nucleic acid fragment of Claim 35 wherein the isolated nucleic acid fragment hybridizes under stringent conditions to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:38.

5 43. The isolated nucleic acid fragment of Claim 35 wherein the isolated nucleic acid fragment is at least 80% similar to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:26.

44. The isolated nucleic acid fragment of Claim 35 wherein the isolated nucleic acid fragment is at least 80% similar to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:28.

10 45. The isolated nucleic acid fragment of Claim 35 wherein the isolated nucleic acid fragment is at least 80% similar to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:30.

46. The isolated nucleic acid fragment of Claim 35 wherein the isolated nucleic acid fragment is at least 80% similar to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:32.

15 47. The isolated nucleic acid fragment of Claim 35 wherein the isolated nucleic acid fragment is at least 80% similar to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:34.

48. The isolated nucleic acid fragment of Claim 35 wherein the isolated nucleic acid fragment is at least 80% similar to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:36.

49. The isolated nucleic acid fragment of Claim 35 wherein the isolated nucleic acid fragment is at least 80% similar to an isolated nucleic acid fragment encoding all or a substantial portion of the amino acid sequence set forth in SEQ ID NO:38.

25 50. The isolated nucleic acid fragment of Claim 35 wherein the nucleotide sequence of the fragment comprises all or a portion of the sequence set forth in a member selected from the group consisting of SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, SEQ ID NO:35 and SEQ ID NO:37.

51. A chimeric gene comprising the nucleic acid fragment of Claim 35 operably linked to suitable regulatory sequences.

52. A transformed host cell comprising the chimeric gene of Claim 51.

53. A tryptophan synthase alpha subunit polypeptide comprising all or a substantial portion of the amino acid sequence set forth in a member selected from the group consisting of SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, SEQ ID NO:36 and SEQ ID NO:38.

54. A method of altering the level of expression of a tryptophan biosynthetic enzyme in a host cell comprising:

- (a) transforming a host cell with the chimeric gene of any of Claims 19, 32 and 51; and

- (b) growing the transformed host cell produced in step (a) under conditions that are suitable for expression of the chimeric gene

wherein expression of the chimeric gene results in production of altered levels of a tryptophan biosynthetic enzyme in the transformed host cell.

5 55. A method of obtaining a nucleic acid fragment encoding all or a substantial portion of the amino acid sequence encoding a tryptophan biosynthetic enzyme comprising:

- 10 (a) probing a cDNA or genomic library with the nucleic acid fragment of any of Claims 1, 22 and 35;
 (b) identifying a DNA clone that hybridizes with the nucleic acid fragment of any of Claims 1, 22 and 35;
 (c) isolating the DNA clone identified in step (b); and
 (d) sequencing the cDNA or genomic fragment that comprises the clone isolated in step (c)

15 wherein the sequenced nucleic acid fragment encodes all or a substantial portion of the amino acid sequence encoding a tryptophan biosynthetic enzyme.

 56. A method of obtaining a nucleic acid fragment encoding a substantial portion of an amino acid sequence encoding a tryptophan biosynthetic enzyme comprising:

- 20 (a) synthesizing an oligonucleotide primer corresponding to a portion of the sequence set forth in any of SEQ ID NOs: 1, 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35 and 37; and
 (b) amplifying a cDNA insert present in a cloning vector using the oligonucleotide primer of step (a) and a primer representing sequences of the cloning vector

25 wherein the amplified nucleic acid fragment encodes a substantial portion of an amino acid sequence encoding a tryptophan biosynthetic enzyme.

 57. The product of the method of Claim 55.

 58. The product of the method of Claim 56.

 59. A method for evaluating at least one compound for its ability to inhibit the activity of a tryptophan biosynthetic enzyme, the method comprising the steps of:

- 30 (a) transforming a host cell with a chimeric gene comprising a nucleic acid fragment encoding a tryptophan biosynthetic enzyme, operably linked to suitable regulatory sequences;
 (b) growing the transformed host cell under conditions that are suitable for expression of the chimeric gene wherein expression of the chimeric
35 gene results in production of the tryptophan biosynthetic enzyme encoded by the operably linked nucleic acid fragment in the transformed host cell;
 (c) optionally purifying the tryptophan biosynthetic enzyme expressed by the transformed host cell;

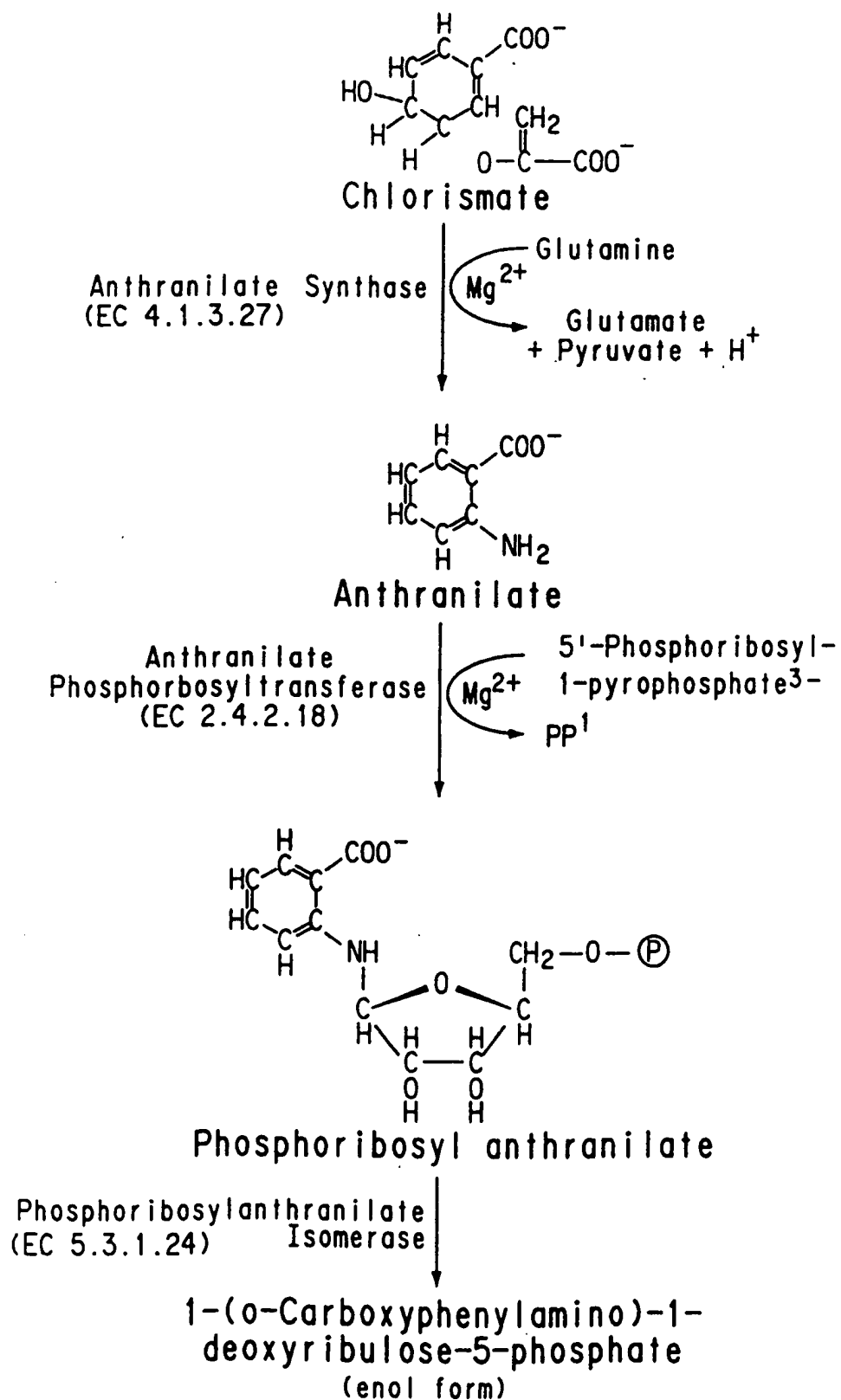
- (d) treating the tryptophan biosynthetic enzyme with a compound to be tested; and
- (e) comparing the activity of the tryptophan biosynthetic enzyme that has been treated with a test compound to the activity of an untreated tryptophan biosynthetic enzyme,

5

thereby selecting compounds with potential for inhibitory activity.

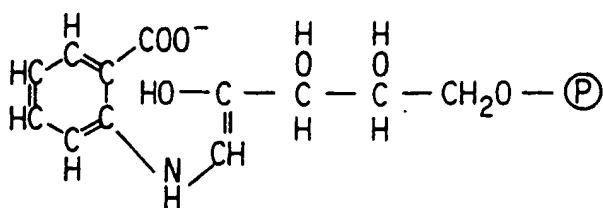
1/9

FIG. 1

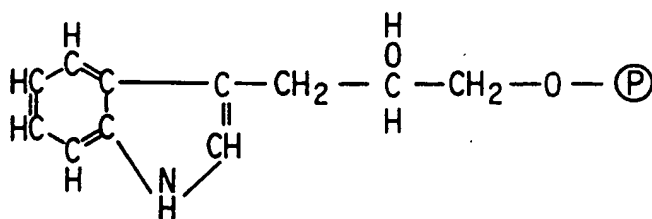


2/9

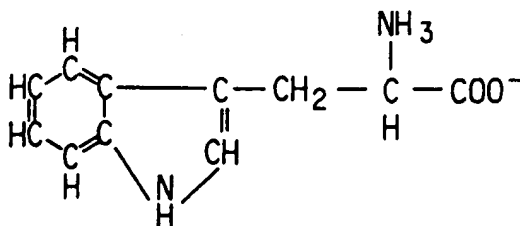
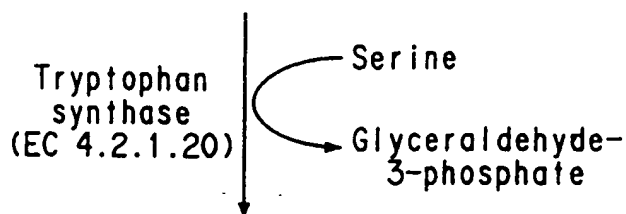
FIG. 1 (CONTINUED)



1-(o-Carboxyphenylamino)-1-deoxyribulose-5-phosphate



Indoleglycerol phosphate



Tryptophan

3/9

Figure 2

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* * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * *
SEQ ID NO:39 MITLNVE TP-PLTRSQLPSTFRVSSAASVNENDRVATSRWRPNLSLTTSSYRLRLTKCA **
SEQ ID NO:02 MESLAATSVFAPSRXAVPARALVRAGTV-----VPTRR-----TSSRSGTSGVKCS 60
1
* * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * *
SEQ ID NO:39 ASASTSASTSASPSPSLVDQSANFHEASKK-GNLIPLYRCIFSDH----LTPVLAYRC ***
SEQ ID NO:02 AAVTPQASPVISRSAAA AAEEDKRRXSRRRGGTGRGTWCPCGSASCRTIYPRARYRC 120
61
* * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * *
SEQ ID NO:39 LVKEDDRDAPSLFESVEPGSQASS-IGRYSVVGAQPAIEIVAKENMVTILDHEGGQRTTE **
SEQ ID NO:02 LVPEDNVXAPSLFESVEQGPQGTNVGRYSMVGAHPVMEIVAKDHKVTIMDHEKSQVTE 180
121
* * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * *
SEQ ID NO:39 QFVEDPMDVPRRIMEGWKPQLIDELPEAFCGGWVGFFSYDTVRYVEKKKLPFFSAPTDDR ***
SEQ ID NO:02 QVVDDPMQIPRTMMEGWHPQQIDELPESEFSGGWVGFFSYDTVRYVEKKKLPFFSAPQDDR 240
181
* * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * *
SEQ ID NO:39 NLPDVHLGLYDDVIVFDHVEKKAFAVIHWVRLDQYSSVAEAYNDGMNRLLENLVS RVHDIVP
SEQ ID NO:02 NLPDVHLGLYDDVIVFDNVEKKVYVIHWVNVDRHASVEEAYQDGRSRLNMLLSKVHNSNV 300
241

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*** ** ** ** **
SEQ ID NO:39 IALALRTMVFQTATRYDTMYSYKDVKRRWIAHLQAGAGIVADSDPADEQRECENKAAA
SEQ ID NO:02 IALSRLTIVFSTAPSHNTMYSYKDAADRNRREWWAHLQAGAGIVADSSPDDEQRECENKAAA

541 600

Figure 3

| | | |
|---------------|---|-----|
| SEQ ID NO: 40 | MAASTLYKSCLLQPKSGST-RRINP-SLVNPLTNPTRVSVLG---KSRRDVFA-KASI | 1 |
| SEQ ID NO: 20 | MACSHIAAAGVSSPAAAAAR-SPAHSASAFARLSTRPRFASAGLSVKGNGAAFLVAAA | 60 |
| SEQ ID NO: 22 | ATPPLFSSDGHRRRAAPPQDPVPRLPGRRGGAEARPSSLRLGVTSGPARTLKQKLVAKS AV | 60 |
| | | |
| SEQ ID NO: 40 | EMAESNSIPSVVNVNSSKQHGPIIIVIDNYDSFTYNLCQYMGEFGCHFEVYRNDELTVVEELK | 61 |
| SEQ ID NO: 20 | GPAAAAAPVADLDGRPATEKQPIIIVIDNYDSFTYNLCQYMGEGLNFEVYRNDELTIEDVR | |
| SEQ ID NO: 22 | SVVEGENAFDGV---KQDTRPIIIVIDNYDSFTYNLCQYMGEVGFANFEVYRNDDITVEEIK | 120 |
| | | |
| SEQ ID NO: 40 | KKNPRGVLI SPGPGTPQDSGISLQTVLELGPLVPLFGVCMGLQCIGEA FGKIVRSPFGV | 121 |
| SEQ ID NO: 20 | RKNPRGILISPGGEPQDSGISLQTVLELGTPIPIFGVCMGLQCIGEA FGKII RAPSGV | |
| SEQ ID NO: 22 | KISPRGILISPGGTPQDSGISLQTVQDLGPSTPLFGVCMGLQCIGEA FGKVVRS PYGV | 180 |
| | | |
| SEQ ID NO: 40 | MHGKSSMVHYDEKGEGLFSGLSNPFIVGRYHSLVIEKDTFPSDELEVTAWTE DGLVMAA | 181 |
| SEQ ID NO: 20 | MHGKSSPVVYDEELGKALFNGLPNPFTAARYHSLVIEQETFPHDAL EATAWTE DGLIMAA | |
| SEQ ID NO: 22 | VHGKGS LVHYEEKLDGTLFSGLPNPFQAGRYHSLVIEKDSFPHDAL EITAWTDDGLIMAA | 240 |

Figure 3.. (Cont..)

| | | | |
|--------------|---------|--|-----|
| SEQ ID NO:40 | * * * * | RHRKYYKHIIQGVQFHPESIIITTEGKTIVRNFIKIVEKKESEKLT | |
| SEQ ID NO:20 | | RHKYYKHIIQGVQFHPESIIITPEGKKIILNFVRFIEELEKQRS | |
| SEQ ID NO:22 | | RTRKSKQYTG..... | 283 |
| | | 241 | |

Figure 4

| | | |
|---------------|---|----|
| SEQ ID NO: 41 |MAIAFKSGVFFLQSPKSIQIGFRHSSPPDSSLSFKRFTPMASLST---- | * |
| SEQ ID NO: 26 |MAFALKAA-----AAGSASFSAAGP-----RRRAAAT-GRVSF- | |
| SEQ ID NO: 28 | ...ARQAQDRKLAIKAAPLRLVRHSVHRPPPPRRLLLTTPQQRQRSEVALCRVEWW | |
| SEQ ID NO: 32 | RTCSSRGHPLRRLHFVAVV-----AVAAAAASLSSAWAALL--RFRGGGG-AGLGLS | |
| | 1 | 60 |

| | | |
|---------------|--|-----------|
| SEQ ID NO: 41 |SSPTLGLADTFTQLKKQKGVAFIPYITAGDPPDLSTTA | * * * * * |
| SEQ ID NO: 26 | ---RS-AAPVVAVRAAAAAAUAEDKRSISGTFaelRQQGKTALIPFITAGDPPDLATTA | |
| SEQ ID NO: 28 | THPRPWPTAALRPASSPPRPSPTSGSKARVGLLAPWWPIPNsAFIPFITAGDPPDLVTTs | |
| SEQ ID NO: 32 | ARDRE.PGRPLMEMEDSGRGVVGAGKRGVAETFSRLREQGKTAfIPFITASDPPDLATTS | |
| | 61 | 120 |

| | | |
|---------------|--|-----------|
| SEQ ID NO: 41 | ** | * * * * * |
| SEQ ID NO: 26 | EALKVLDACGSDIIElGVpYSDPLADGPVlQAaATRSlerGtNLDsileMlDKVVPQlSC | |
| SEQ ID NO: 28 | KALRILDACGSDVIElGVpYSDPLADGPVlQASATRALAKGtTFEDVlSMVKGVlPDLSC | |
| SEQ ID NO: 32 | KALKILNSCGSDVIEVGVpYSDPLADGPVlQASATRALKKGtTLDsVlEMLKGvTPeLSC | |
| | KALKILDSCGSDVIElGVpYSDPLADGPVlQAaATRALKKGATFDsVlAMlKGvIPELSC | 180 |
| | 121 | |

Figure 4 (Cont.)

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*      *      *      *      *      *      *      *      *      *      *      *      *      *      *
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SEQ ID NO: 26      PVALFTYYPILKRGVPNFMMSIVKEAGVHGLVVPDVPLEETDVLRSEAAKNNLELVLLTT
SEQ ID NO: 28      PIVLFTYYPILKRGVGNFMSTIKQAGIHGLVVPDLPLEETALLRSEAIMHNIELVLLTT
SEQ ID NO: 32      PIVIFTYYPILKRGVSNFMAIKQAGVHGLVVPDLPLEETALLRNEAVMHGIELVLLTT      240
181

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SEQUENCE LISTING

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<120> TRYPTOPHAN BIOSYNTHETIC ENZYMES

<130> BB-1150

<140>

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<150> 60/079,386

<151> 1998-03-26

<160> 41

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 <213> Zea mays

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 35 40 45
 Ala Val Thr Pro Gln Ala Ser Pro Val Ile Ser Arg Ser Ala Ala Ala
 50 55 60
 Ala Lys Ala Ala Glu Glu Asp Lys Arg Arg Xaa Ser Arg Arg Arg Arg
 65 70 75 80
 Gly Gly Thr Gly Arg Gly Thr Trp Cys Pro Cys Gly Ser Ala Ser Cys
 85 90 95
 Arg Thr Ile Tyr Pro Arg Ala Arg Tyr Arg Cys Leu Val Pro Glu Asp
 100 105 110
 Asn Val Xaa Ala Pro Ser Phe Leu Phe Glu Ser Val Glu Gln Gly Pro
 115 120 125
 Gln Gly Thr Thr Asn Val Gly Arg Tyr Ser Met Val Gly Ala His Pro
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 Val Met Glu Ile Val Ala Lys Asp His Lys Val Thr Ile Met Asp His
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 Glu Lys Ser Gln Val Thr Glu Gln Val Val Asp Asp Pro Met Gln Ile
 165 170 175
 Pro Arg Thr Met Met Glu Gly Trp His Pro Gln Gln Ile Asp Glu Leu
 180 185 190

Pro Glu Ser Phe Ser Gly Gly Trp Val Gly Phe Phe Ser Tyr Asp Thr
 195 200 205
 Val Arg Tyr Val Glu Lys Lys Lys Leu Pro Phe Ser Ser Ala Pro Gln
 210 215 220
 Asp Asp Arg Asn Leu Pro Asp Val His Leu Gly Leu Tyr Asp Asp Val
 225 230 235 240
 Leu Val Phe Asp Asn Val Glu Lys Lys Val Tyr Val Ile His Trp Val
 245 250 255
 Asn Val Asp Arg His Ala Ser Val Glu Glu Ala Tyr Gln Asp Gly Arg
 260 265 270
 Ser Arg Leu Asn Met Leu Leu Ser Lys Val His Asn Ser Asn Val Pro
 275 280 285
 Thr Leu Ser Pro Gly Phe Val Lys Leu His Thr Arg Lys Phe Gly Thr
 290 295 300
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 325 330 335
 Leu Ser Gln Arg Phe Glu Arg Arg Thr Tyr Ala Asn Pro Phe Glu Val
 340 345 350
 Tyr Arg Ala Leu Arg Ile Val Asn Pro Ser Pro Tyr Met Ala Tyr Val
 355 360 365
 Gln Ala Arg Gly Cys Val Leu Val Ala Ser Ser Pro Glu Ile Leu Thr
 370 375 380
 Arg Val Ser Lys Gly Lys Ile Ile Asn Arg Pro Leu Ala Gly Thr Val
 385 390 395 400
 Arg Arg Gly Lys Thr Glu Lys Glu Asp Gln Met Gln Glu Gln Gln Leu
 405 410 415
 Leu Ser Asp Glu Lys Gln Cys Ala Glu His Ile Met Leu Val Asp Leu
 420 425 430
 Gly Arg Asn Asp Val Gly Lys Val Ser Lys Pro Gly Ser Val Lys Val
 435 440 445
 Glu Lys Leu Met Asn Ile Glu Arg Tyr Ser His Val Met His Ile Ser
 450 455 460
 Ser Thr Val Ser Gly Gln Leu Asp Asp His Leu Gln Ser Trp Asp Ala
 465 470 475 480
 Leu Arg Ala Ala Leu Pro Val Gly Thr Val Ser Gly Ala Pro Lys Val
 485 490 495
 Lys Ala Met Glu Leu Ile Asp Lys Leu Glu Val Thr Arg Arg Gly Pro
 500 505 510
 Tyr Ser Gly Gly Leu Gly Gly Ile Ser Phe Asp Gly Asp Met Gln Ile
 515 520 525
 Ala Leu Ser Leu Arg Thr Ile Val Phe Ser Thr Ala Pro Ser His Asn
 530 535 540

Thr Met Tyr Ser Tyr Lys Asp Ala Asp Arg Arg Arg Glu Trp Val Ala
545 550 555 560

His Leu Gln Ala Gly Ala Gly Ile Val Ala Asp Ser Ser Pro Asp Asp
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Glu Gln Arg Glu Cys Glu Asn Lys Ala Ala Ala Leu Ala Arg Ala Ile
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<213> Zea mays

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 35 40 45
 Ser Tyr Asp Thr Val Arg Tyr Val Glu Thr Lys Lys Leu Pro Phe Ser
 50 55 60
 Lys Ala Pro His Asp Asp Arg Asn Leu Pro Asp Ile His Leu Gly Leu
 65 70 75 80
 Tyr Ser Asp Val Ile Val Phe Asp His Val Glu Lys Lys Thr His Val
 85 90 95
 Ile His Trp Val Arg Thr His Cys Tyr Arg Ser Val Asp Glu Ala Tyr
 100 105 110
 Glu Asp Gly Arg Asn Arg Leu Glu Ala Leu Leu Ser Arg Leu His Cys
 115 120 125
 Leu Asn Val Pro Thr Leu Ser Ser Gly Ser Ile Lys Leu Asn Val Glu
 130 135 140
 Asn Phe Gly Pro Val Met Gln Lys Ser Thr Met Ser Ser Glu Glu Tyr
 145 150 155 160
 Lys Asn Ile Val Val Gln Ala Lys Glu His Ile Leu Ala Gly Asp Ile
 165 170 175
 Phe Gln Val Val Leu Ser Gln Arg Phe Glu Arg Arg Thr Phe Ala Asp
 180 185 190
 Pro Phe Glu Ile Tyr Arg Ala Leu Arg Ile Val Asn Pro Ser Pro Tyr
 195 200 205
 Met Ala Tyr Leu Gln Ala Arg Gly Cys Ile Leu Val Ala Ser Ser Pro
 210 215 220
 Glu Ile Leu Thr Arg Val Gln Lys Arg Thr Ile Ile Asn Arg Pro Leu
 225 230 235 240
 Ala Gly Thr Ile Arg Arg Gly Lys Thr Lys Ala Glu Asp Lys Thr Leu
 245 250 255
 Glu Gln Leu Leu Leu Ser Asp Glu Lys Gln Cys Ala Glu His Ile Met
 260 265 270
 Leu Val Asp Leu Gly Arg Asn Asp Val Gly Lys Val Ser Lys Pro Gly
 275 280 285
 Ser Val Lys Val Glu Lys Leu Met Asn Ile Glu Arg Tyr Ser His Val
 290 295 300
 Met His Ile Ser Ser Thr Val Thr Gly Glu Leu Arg Asp Asp Leu Thr
 305 310 315 320
 Cys Trp Asp Ala Leu Arg Ala Ala Leu Pro Val Gly Thr Val Ser Gly
 325 330 335
 Ala Pro Lys Val Arg Ala Met Glu Leu Ile Asp Gln Leu Glu Val Ser
 340 345 350
 Met Arg Gly Pro Tyr Ser Gly Gly Phe Gly Gly Ile Ser Phe Arg Gly
 355 360 365
 Asp Met Asp Ile Ala Leu Ala Leu Arg Thr Ile Val Phe Pro Thr Ala
 370 375 380

Ser Arg Phe Asp Thr Met Tyr Ser Tyr Thr Asp Ser Lys Ser Arg Gln
385 390 395 400

Glu Trp Val Ala His Leu Gln Ala Gly Ala Gly Ile Val Ala Asp Ser
405 410 415

Lys Pro Asp Asp Glu His Gln Glu Cys Ile Asn Lys Ala Ala Gly Val
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Ala Arg Ala Ile Asp Leu Ala Glu Ser Thr Phe Leu Glu Glu
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Xaa His Thr Val Ala Gly Leu Gly Gly
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 35 40 45
 Val Ile His Trp Val Arg Val Asp Arg Tyr Ser Ser Ala Glu Glu Ala
 50 55 60
 Phe Glu Asp Gly Arg Asn Arg Leu Glu Thr Leu Xaa Ser Arg Val His
 65 70 75 80
 Asp Ile Ile Thr Pro Arg Leu Pro Thr Gly Ser Ile Lys Leu Tyr Thr
 85 90 95
 Arg Leu Phe Gly Pro Lys Leu Glu Met Ser Xaa Met Thr Asn Glu Glu
 100 105 110
 Xaa Lys Arg Ala Val Leu Lys Ala Lys Glu His
 115 120

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 <212> DNA
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 aagaagggga acgtcattcc tctcttcgc tgcataatct ccgatcacct cactccggtg 240
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 <213> Glycine max

<220>
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 Arg Ser Thr Leu Lys Cys Cys Ala Gln Ser Pro Ser Pro Ser Leu Val
 35 40 45
 Asp Asn Ala Gln Lys Phe Leu Glu Ala Ser Lys Lys Gly Asn Val Ile
 50 55 60
 Pro Leu Phe Arg Cys Ile Phe Ser Asp His Leu Thr Pro Val Leu Ala
 65 70 75 80
 Tyr Arg Cys Leu Val Lys Glu Asp Glu Arg Asp Ala Pro Ser Phe Leu
 85 90 95
 Phe Glu Ser Val Glu Pro Gly Gln Ile Ser Ser Ile Gly Arg Tyr Ser
 100 105 110
 Val Val Gly Ala Gln Ala Val Xaa Trp Glu Ile Val Ala Lys Glu Asn
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 Val Gly
 130

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 <212> DNA
 <213> Triticum aestivum

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 accttcctga tgttcacttg gggctttacg atgatgttct tgtcttcgac aatgttgaga 180
 agaaagtata tgtcatccat tgggtaagtg tggaccggca tgcattccacc gaggaagcat 240
 acaaagatgg cagggtcccg ttgaagcggg tgctttctaa agttcacaat gcaaagtgtcc 300
 ccaagctctc tccaggattt gtgaagctac atactcggca gtttggtact cccttgaaca 360
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 <212> PRT
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 20 25 30
 Gly Ala Pro His Asp Asp Arg Asn Leu Pro Asp Val His Leu Gly Leu
 35 40 45
 Tyr Asp Asp Val Leu Val Phe Asp Asn Val Glu Lys Lys Val Tyr Val
 50 55 60
 Ile His Trp Val Ser Val Asp Arg His Ala Ser Thr Glu Glu Ala Tyr
 65 70 75 80
 Lys Asp Gly Arg Ser Arg Leu Lys Arg Leu Leu Ser Lys Val His Asn
 85 90 95
 Ala Asn Val Pro Lys Leu Ser Pro Gly Phe Val Lys Leu His Thr Arg
 100 105 110
 Gln Phe Gly Thr Pro Leu Asn Lys Ser Thr Met Thr Ser Asp Glu Tyr
 115 120 125
 Lys Ser Ala Val Met Gln Ala Lys Glu His Ile Leu Gly Gly Asn Ile
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 Phe Gln Ile Val Leu Ser
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<210> 13
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 <212> DNA
 <213> Triticum aestivum

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 ctcatagagg ccccgagagt ggggtgctca tcttcaggct ggtgcgggca ttgttgctga 180
 tagtatccca gacgatgagc aaaaagaatt tgagaataag gcggctgccc tagctcgggc 240
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 ttttagttgt tcatcatttt tcacccagtt cattttggaa agttgttcat cgttttttca 360
 ccgagttcat attggggaaa aaaagcaata ccgttttgtt gtcctttgaa atgaataaat 420
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 aaaaaaaaaa aaaaaaaaaa aata 504

<210> 14
 <211> 92
 <212> PRT
 <213> Triticum aestivum

<400> 14
 Pro Asn Ser Gly Gly Leu Gly Gly Ile Ser Phe Asp Gly Asp Met Leu
 1 5 10 15
 Ile Ala Leu Ala Leu Arg Thr Ile Val Phe Ser Thr Ala Pro Ser Pro
 20 25 30
 Asn Arg Met Tyr Ser Tyr Lys Ser Ser Asp Arg Pro Arg Glu Trp Val
 35 40 45
 Ala His Leu Gln Ala Gly Ala Gly Ile Val Ala Asp Ser Ile Pro Asp
 50 55 60
 Asp Glu Gln Lys Glu Phe Glu Asn Lys Ala Ala Ala Leu Ala Arg Ala
 65 70 75 80
 Ile Asp Leu Ala Glu Ser Ala Phe Leu Asp Lys Glu
 85 90

<210> 15
 <211> 535
 <212> DNA
 <213> Triticum aestivum

<220>
 <221> unsure
 <222> (483)

<220>
 <221> unsure
 <222> (511)

<220>
 <221> unsure
 <222> (520)..(521)

<400> 15
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 agagcagtgt cagaccgttg ttgagcagga agcggaagga cagttcaatc cgcaatggaa 180
 tccctagccg ccgccacgtt ctgcgccctcg cgcctcgctg cccaccccg cccgcccgcg 240
 gctgcggcag cgggcagatc gagggcggtg gcggcaacga ggaggaggag gagcagcagc 300
 ggccctgaggt gctcgtcggc gagcgcgacc ccggtgatca atgggagcgc cgccgcgaag 360
 gcggaggagg aggacaggag gcgcttcttt gaggcggcgc cacgggggac cggcaagggc 420
 aacctggtgc ccgtgtggga gtgcacgtg tgcgattacc tcaccccccgt gctcgcctaa 480
 cgntgcctcg ttcccgaagg acgacattgg ncaacccan nttcctcttc caagg 535

<210> 16
 <211> 108
 <212> PRT
 <213> Triticum aestivum

<220>
 <221> UNSURE
 <222> (102)

<400> 16
 Met Glu Ser Leu Ala Ala Ala Thr Phe Ser Pro Ser Arg Leu Ala Ala
 1 5 10 15

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<400>      18
His Glu Glu Ile Lys Lys Ile Ser Pro Arg Gly Ile Leu Ile Ser Pro
   1              5              10              15

Gly Pro Gly Thr Pro Gln Asp Ser Gly Ile Ser Leu Gln Thr Val Thr
          20              25              30

Glu Leu Gly Pro Ser Ile Pro Leu Phe Gly Val Cys Met Gly Leu Gln
          35              40              45

Cys Ile Gly Glu Ala Phe Gly Gly Lys Val Val Arg Ser Pro Tyr Gly
          50              55              60

Val Val His Gly Lys Gly Ser Leu Val His Tyr Asp Glu Lys Leu Asp
   65              70              75              80

Gly Thr Leu Phe Tyr Asp Ile Pro Asn Pro Phe Gln Ala Gly Arg Tyr
          85              90              95

His Ser Leu Val Ile Glu Lys Asp Ser Phe Pro His Asp Thr Leu Glu
          100              105              110

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Ile Val Ala Trp Thr Asp Asp Gly Leu Ile Met Ala Ala Arg His Arg
115 120 125

Ile Tyr Lys His Ile Gln Gly Val Gln Phe His Pro Glu Ser Ile Ile
130 135 140

Thr Thr Glu Gly Arg Leu Met Val Asn Asn Phe Ile Lys Ile Ile Glu
145 150 155 160

Gly Tyr Glu Ala Ser Asn Cys Ser Pro
165

<210> 19
<211> 1133
<212> DNA
<213> Zea mays

<220>
<221> unsure
<222> (13)

<400> 19
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cgacgcctcg ttctcgcgagc gctggcttgt cgggttaagg aaacggagcg gcgttcccgt 180
tggtcgccgc cgcggggcgc gccgcggcgg caccggtggc cgacctggac ggccgcccgg 240
caacggagaa gcagcccatc atcgatcatc acaactacga cagcttcaca tacaacctct 300
gccagtatat gggggagctt ggattgaact tcgaagtata ccgcaatgat gaactgacca 360
tagaagatgt gagaaggaag aacccaaggg gaatacttat ttctccagga cctgggtgaac 420
cacaagattc gggaatatca ttgcagactg ttcttgaatt aggcccaacc atcccaattt 480
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aactggagaa gcagcggttc taggggaggt agacgcgaca ggtggtttca ttcatagat 900
cagacggaag cagagacaag gcgcttaaa gctgcgccagg ttccgggggt ggcagttgaa 960
gcatagctag gaaacagctt tccctctctt aattcggttg gtctgtggtg acataatctg 1020
tgtgcggacc gaatttagaa taaagtacag gcgtccgtct gatcaataa gcaccggtgt 1080
tggtgctcta tagatgaata tatccggtga ttttcggcaa aaaaaaaaaa aaa 1133

<210> 20
<211> 281
<212> PRT
<213> Zea mays

<400> 20
Met Ala Cys Ser His Ile Ala Ala Ala Gly Val Ser Ser Pro Ala Ala
1 5 10 15
Ala Ala Ala Arg Ser Pro Ala His Ser Pro Ala Ser Ala Phe Ala Arg
20 25 30
Leu Arg Ser Thr Pro Arg Phe Ala Ser Ala Gly Leu Ser Val Lys Gly
35 40 45
Asn Gly Ala Ala Phe Pro Leu Val Ala Ala Ala Gly Pro Ala Ala Ala
50 55 60
Ala Pro Val Ala Asp Leu Asp Gly Arg Pro Ala Thr Glu Lys Gln Pro
65 70 75 80
Ile Ile Val Ile Asp Asn Tyr Asp Ser Phe Thr Tyr Asn Leu Cys Gln
85 90 95

Tyr Met Gly Glu Leu Gly Leu Asn Phe Glu Val Tyr Arg Asn Asp Glu
 100 105 110
 Leu Thr Ile Glu Asp Val Arg Arg Lys Asn Pro Arg Gly Ile Leu Ile
 115 120 125
 Ser Pro Gly Pro Gly Glu Pro Gln Asp Ser Gly Ile Ser Leu Gln Thr
 130 135 140
 Val Leu Glu Leu Gly Pro Thr Ile Pro Ile Phe Gly Val Cys Met Gly
 145 150 155 160
 Leu Gln Cys Ile Gly Glu Ala Phe Gly Gly Lys Ile Ile Arg Ala Pro
 165 170 175
 Ser Gly Val Met His Gly Lys Ser Ser Pro Val Tyr Tyr Asp Glu Glu
 180 185 190
 Leu Gly Lys Ala Leu Phe Asn Gly Leu Pro Asn Pro Phe Thr Ala Ala
 195 200 205
 Arg Tyr His Ser Leu Val Ile Glu Gln Glu Thr Phe Pro His Asp Ala
 210 215 220
 Leu Glu Ala Thr Ala Trp Thr Glu Asp Gly Leu Ile Met Ala Ala Arg
 225 230 235 240
 His Lys Lys Tyr Lys His Ile Gln Gly Val Gln Phe His Pro Glu Ser
 245 250 255
 Ile Ile Thr Pro Glu Gly Lys Lys Ile Ile Leu Asn Phe Val Arg Phe
 260 265 270
 Ile Glu Glu Leu Glu Lys Gln Arg Ser
 275 280

<210> 21
 <211> 856
 <212> DNA
 <213> Oryza sativa

<220>
 <221> unsure
 <222> (811)

<220>
 <221> unsure
 <222> (816)..(816)

<220>
 <221> unsure
 <222> (831)

<220>
 <221> unsure
 <222> (836)

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 tccccggccg tcgaggaggc ggggaggcgc gccctccag tctccgatta ggagttacta 180
 gtggaccgc aagaactctg aagcaaaagc ttgttgctaa gagtgctgtt tctgtggtgg 240
 aaggtgaaaa cgcatattgat ggagtaaagc aagatactag accaatcata gttatagata 300
 actacgatag cttcacgtat aatttatgcc agtacatggg tgagggtggga gctaactttg 360
 aggtgtaccg caatgattga taccaccgtg gaagaaatta agaagatttc tcctagagga 420
 atactcatct cccctggccc tggcacacct caagattcag gaatatcatt gcaaacagtt 480

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caagatcttg gaccttctac acctttgttt ggggtttgca tgggtttgca gtgtattggg 540
gaggcatttg gaggggaaggt tggtcgttct ccttatggag ttgtgcatgg gaaaggatcc 600
cttgttcact atgaggagaa acttgatgga acactgtttt ctggtctccc aaaccattc 660
caagcgggaa gataccacag ccttgtaatt gagaaggata gcttcccaca tgatgccctg 720
gaaattactg cttggacaga cgatgggctg atcatggctg ctcgcacaag gaagtccaaa 780
caatatacag ggtggtgcag tccatccaag ngagcnatca taacaccaga ngggangctc 840
aatggtccaa gaattt                                     856

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<210> 22
 <211> 247
 <212> PRT
 <213> *Oryza sativa*

<400> 22
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 20 25 30
 Ala Arg Pro Ser Ser Leu Arg Leu Gly Val Thr Ser Gly Pro Ala Arg
 35 40 45
 Thr Leu Lys Gln Lys Leu Val Ala Lys Ser Ala Val Ser Val Val Glu
 50 55 60
 Gly Glu Asn Ala Phe Asp Gly Val Lys Gln Asp Thr Arg Pro Ile Ile
 65 70 75 80
 Val Ile Asp Asn Tyr Asp Ser Phe Thr Tyr Asn Leu Cys Gln Tyr Met
 85 90 95
 Gly Glu Val Gly Ala Asn Phe Glu Val Tyr Arg Asn Asp Asp Ile Thr
 100 105 110
 Val Glu Glu Ile Lys Lys Ile Ser Pro Arg Gly Ile Leu Ile Ser Pro
 115 120 125
 Gly Pro Gly Thr Pro Gln Asp Ser Gly Ile Ser Leu Gln Thr Val Gln
 130 135 140
 Asp Leu Gly Pro Ser Thr Pro Leu Phe Gly Val Cys Met Gly Leu Gln
 145 150 155 160
 Cys Ile Gly Glu Ala Phe Gly Gly Lys Val Val Arg Ser Pro Tyr Gly
 165 170 175
 Val Val His Gly Lys Gly Ser Leu Val His Tyr Glu Glu Lys Leu Asp
 180 185 190
 Gly Thr Leu Phe Ser Gly Leu Pro Asn Pro Phe Gln Ala Gly Arg Tyr
 195 200 205
 His Ser Leu Val Ile Glu Lys Asp Ser Phe Pro His Asp Ala Leu Glu
 210 215 220
 Ile Thr Ala Trp Thr Asp Asp Gly Leu Ile Met Ala Ala Arg Thr Arg
 225 230 235 240
 Lys Ser Lys Gln Tyr Thr Gly
 245

<210> 23
 <211> 468
 <212> DNA
 <213> *Triticum aestivum*

<220>

<221> unsure

<222> (7)

<400> 23

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ggaacncttc ccgcacgttg aactggaagt cacggcatgg actgaagatg gacttgtcat 60
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atcatcaccc ctgaaggcaa gaaaatcatc ctcaactttg cgagatatgt cgaggagttt 180
gagaagcaga cctccgaggg gaagtagagg attcaggaag cagagcagtt agaggaagta 240
gagactattg ttgagagatc aggcaggcag gaaggggctt aaaagccaag cagatttagg 300
ggaaatgaat aagggaaaaa gagcttttcc attgccattc gtttggttca ttgtaataa 360
tcattgtgga tctagatccc gtatgaactt cattcgggtca attatgttcg tcgtgggtgct 420
ctttctcaaa taataaatcg gtggaagcaa aaaaaaaaaa aaaactcg 468

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<210> 24

<211> 67

<212> PRT

<213> Triticum aestivum

<400> 24

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Glu Pro Phe Pro His Val Glu Leu Glu Val Thr Ala Trp Thr Glu Asp
  1                      5                      10                      15

```

```

Gly Leu Val Met Ala Ala Arg His Lys Lys Tyr Thr His Ile Gln Gly
                20                      25                      30

```

```

Val Gln Phe His Pro Glu Ser Ile Ile Thr Pro Glu Gly Lys Lys Ile
        35                      40                      45

```

```

Ile Leu Asn Phe Ala Arg Tyr Val Glu Glu Phe Glu Lys Gln Thr Ser
        50                      55                      60

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Glu Gly Lys
  65

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<210> 25

<211> 1181

<212> DNA

<213> Zea mays

<400> 25

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aggccgcgcg cgcgggggtcg gcgtcggtct ccgccgcggg gccgagggcg cgcgccgcgg 120
cgactggtag ggtctcggtc cggagcgccg cgcccggtgt cgccgtcagg gccgcggcgg 180
cgcgggccgc cgctgtggcg gaggacaagc gcagcatctc cggcaccttc gccgagctca 240
ggcagcaggg gaagactgcc ttgattccct tcatcactgc tggagacctt gacttggcca 300
ccacagcaaa agcactcagg atccttgatg cgtgtggttc agacgtgatc gaactgggtg 360
tgccttactc cgatccattg gctgacggcc ctgttattca ggccctcccg acgcgcgctc 420
tggcaaaggg caccacattt gaggatgtca tctccatggt aaaggggggtg atacctgatc 480
tgtcctgccc tgtagcgctt ttcacatatt acaaccgat cctgaagcgt ggtgtcccca 540
acttcatgag tattgttaaa gaagctgggg tacacggtct tgtgttacca gatgttcctt 600
tggaagagac agatgtcttg aggagtgagg ctgccaaaaa caacctagag ctggtgctat 660
tgacaacacc gactacacca aatgaaagaa tggagaaaaa tgcacaggct tcacaaggat 720
ttatttatct tgtaagcact gttggagtta ctggtacacc tgcaaatgta agcggaagg 780
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gtgtgtcgac tccagagcat gtccggcaga ttgcaggatg gggcgcatg ggtgtgatta 900
tcggtagcgc agtgatgaag acgttgaggg aagctgcttc tccagaagaa ggattgaaga 960
agctagaaga gttcgccaag aacctgaagg ccgcattgcc gtgaaccaac cattttcgtt 1020
agcaaaagaa ttaagatttt tttttccttt caatcttggc tttcagggtt gttggttgtt 1080
gtgtagtga aataattgtg tccctatggt gaatgctcaa atgaataaat caagatccca 1140
taaaaaaaaa aaaaaaaaaa tcgagggggg gcccgtagcc a 1181

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<210> 26

<211> 319

<212> PRT
<213> Zea mays

<400> 26

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Met Ala Phe Ala Leu Lys Ala Ala Ala Ala Gly Ser Ala Ser Phe Ser
 1           5           10           15

Ala Ala Gly Pro Arg Arg Arg Ala Ala Ala Thr Gly Arg Val Ser Phe
 20           25           30

Arg Ser Ala Ala Pro Val Val Ala Val Arg Ala Ala Ala Ala Ala
 35           40           45

Ala Ala Val Ala Glu Asp Lys Arg Ser Ile Ser Gly Thr Phe Ala Glu
 50           55           60

Leu Arg Gln Gln Gly Lys Thr Ala Leu Ile Pro Phe Ile Thr Ala Gly
 65           70           75           80

Asp Pro Asp Leu Ala Thr Thr Ala Lys Ala Leu Arg Ile Leu Asp Ala
 85           90           95

Cys Gly Ser Asp Val Ile Glu Leu Gly Val Pro Tyr Ser Asp Pro Leu
100           105           110

Ala Asp Gly Pro Val Ile Gln Ala Ser Ala Thr Arg Ala Leu Ala Lys
115           120           125

Gly Thr Thr Phe Glu Asp Val Ile Ser Met Val Lys Gly Val Ile Pro
130           135           140

Asp Leu Ser Cys Pro Val Ala Leu Phe Thr Tyr Tyr Asn Pro Ile Leu
145           150           155           160

Lys Arg Gly Val Pro Asn Phe Met Ser Ile Val Lys Glu Ala Gly Val
165           170           175

His Gly Leu Val Val Pro Asp Val Pro Leu Glu Glu Thr Asp Val Leu
180           185           190

Arg Ser Glu Ala Ala Lys Asn Asn Leu Glu Leu Val Leu Leu Thr Thr
195           200           205

Pro Thr Thr Pro Asn Glu Arg Met Glu Lys Ile Ala Gln Ala Ser Gln
210           215           220

Gly Phe Ile Tyr Leu Val Ser Thr Val Gly Val Thr Gly Thr Pro Ala
225           230           235           240

Asn Val Ser Gly Lys Val Gln Ser Leu Leu Gln Asp Ile Lys Asn Val
245           250           255

Thr Glu Lys Pro Val Ala Val Gly Phe Gly Val Ser Thr Pro Glu His
260           265           270

Val Arg Gln Ile Ala Gly Trp Gly Ala Asp Gly Val Ile Ile Gly Ser
275           280           285

Ala Val Met Lys Thr Leu Glu Glu Ala Ala Ser Pro Glu Glu Gly Leu
290           295           300

Lys Lys Leu Glu Glu Phe Ala Lys Asn Leu Lys Ala Ala Leu Pro
305           310           315

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<210> 27
<211> 1243

<212> DNA
<213> Zea mays

<400> 27
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ccacaaaggc agcgctcggg ggtggccctc tgtcgggtgg agtgggtggac acatccgcgc 180
ccatggccaa cggcgggcgt gcggccggca agctcaccgt cggcgagacc ttctccaacc 240
tcagggagca aggcaagggt tggattgctg gcgccgtggt ggccgatacc caatagtgcg 300
ttcatcccgt tcataactgc tggatgacct gacctggtaa ccacatcgaa agcattgaag 360
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gctgatgggc cagttattca ggcttctgca acacgggcgc ttaagaaagg cactacactt 480
gattctgtca tagagatgct gaaggggggt acacctgagc tttcttgccc cattgttctt 540
ttcacatact ataatccaat tctgaaacgt ggtgtgggaa acttcatgtc tactatcaaa 600
caagctggca tacatggact tgtagtgcct gatcttcctt tggaggagac agcgttgctg 660
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taaaactgagt tagtgcgaaa aaaaagataa gccttaaaaa aaa 1243

<210> 28
<211> 347
<212> PRT
<213> Zea mays

<400> 28
Ala Arg Ala Gln Ala Asp Arg Lys Leu Ala Ile Lys Ala Ala Pro Leu
1 5 10 15
Arg Leu Arg Val Arg His Ser Val His Arg Pro Pro Pro Pro Arg Pro
20 25 30
Pro Pro Arg Arg Leu Leu Leu Thr Pro Gln Arg Gln Arg Ser Glu Val
35 40 45
Ala Leu Cys Arg Val Glu Trp Trp Thr His Pro Arg Pro Trp Pro Thr
50 55 60
Ala Ala Leu Arg Pro Ala Ser Ser Pro Ser Pro Arg Pro Ser Pro Thr
65 70 75 80
Ser Gly Ser Lys Ala Arg Val Gly Leu Leu Ala Pro Trp Trp Pro Ile
85 90 95
Pro Asn Ser Ala Phe Ile Pro Phe Ile Thr Ala Gly Asp Pro Asp Leu
100 105 110
Val Thr Thr Ser Lys Ala Leu Lys Ile Leu Asn Ser Cys Gly Ser Asp
115 120 125
Val Ile Glu Val Gly Val Pro Tyr Ser Asp Pro Leu Ala Asp Gly Pro
130 135 140
Val Ile Gln Ala Ser Ala Thr Arg Ala Leu Lys Lys Gly Thr Thr Leu
145 150 155 160
Asp Ser Val Ile Glu Met Leu Lys Gly Val Thr Pro Glu Leu Ser Cys
165 170 175

Pro Ile Val Leu Phe Thr Tyr Tyr Asn Pro Ile Leu Lys Arg Gly Val
 180 185 190

Gly Asn Phe Met Ser Thr Ile Lys Gln Ala Gly Ile His Gly Leu Val
 195 200 205

Val Pro Asp Leu Pro Leu Glu Glu Thr Ala Leu Leu Arg Ser Glu Ala
 210 215 220

Ile Met His Asn Ile Glu Leu Val Leu Leu Thr Thr Pro Thr Thr Pro
 225 230 235 240

Thr Asp Arg Met Lys Gly Ile Ala Gln Ala Ser Glu Gly Phe Leu Tyr
 245 250 255

Leu Val Ser Ala Val Gly Val Thr Gly Ala Pro Ser Asn Val Asn Leu
 260 265 270

Arg Val Glu His Leu Leu Arg Glu Ser Lys Lys Val Thr Asp Lys Pro
 275 280 285

Val Ala Val Gly Phe Gly Val Ser Thr Pro Glu His Val Lys Gln Ile
 290 295 300

Val Gly Trp Gly Ala Asp Gly Val Ile Val Gly Ser Ala Ile Val Lys
 305 310 315 320

Gln Leu Cys Glu Ala Ala Thr Pro Glu Glu Gly Leu Glu Arg Leu Glu
 325 330 335

Glu Tyr Ala Arg Ser Met Lys Ala Ala Met Pro
 340 345

<210> 29
 <211> 831
 <212> DNA
 <213> Oryza sativa

<400> 29
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 ttcatgactg ttgtaaaaga agccgggtgct catggtcttg tggtagctga tgttcctctg 120
 gaagagacaa atattttgag gagtgaagct gctaagaaca accttgagct ggtgctactg 180
 acaacaccaa ctacaccaac agaaagaatg gagaaaatta caaaagcttc tgaaggattt 240
 atttatcttg taagcactgt tggagttaca ggtgcacgtg caaatgtcag tggcaagggtg 300
 caatctcttc tccaggatat caagcaggctc acggacaaaag cttgtggctg ttgggttcgg 360
 tatatcgact ccagagcatg tgaagcagat tgcgggatgg ggcgcagatg gtgtgatcat 420
 tgggagcgt atggtgaggc aattgggcga agctgcttca cccgaagaag gattgaagaa 480
 gctagaagag ctagccaaga gcctgaaggc tgcattgccc tgaacaatc agttccatga 540
 ggaaagaaaa tacagtccat tttttttag tagcagtttga agcatagtat tgggagcatg 600
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 gttagttgct tgaacgaata aatcaaggta agaatagtac taaatgttgc gaaaagggcc 720
 taacactggt ttcgctttgt cggttggtgca aatatcgaag agcggatata aacggagaaa 780
 ataatgtcgt aattcaccta aaaaaaaaaa aaaaaaaaaa aaaaaaaaaa a 831

<210> 30
 <211> 179
 <212> PRT
 <213> Oryza sativa

<400> 30
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 1 5 10 15

Gly Ile Ala Asn Phe Met Thr Val Val Lys Glu Ala Gly Val His Gly
 20 25 30

Leu Val Val Pro Asp Val Pro Leu Glu Glu Thr Asn Ile Leu Arg Ser
 35 40 45
 Glu Ala Ala Lys Asn Asn Leu Glu Leu Val Leu Leu Thr Thr Pro Thr
 50 55 60
 Thr Pro Thr Glu Arg Met Glu Lys Ile Thr Lys Ala Ser Glu Gly Phe
 65 70 75 80
 Ile Tyr Leu Val Ser Thr Val Gly Val Thr Gly Ala Arg Ala Asn Val
 85 90 95
 Ser Gly Lys Val Gln Ser Leu Leu Gln Asp Ile Lys Gln Val Thr Asp
 100 105 110
 Lys Ala Cys Gly Cys Trp Val Arg Tyr Ile Asp Ser Arg Ala Cys Glu
 115 120 125
 Ala Asp Cys Gly Met Gly Arg Arg Trp Cys Asp His Trp Glu Arg Tyr
 130 135 140
 Gly Glu Ala Ile Gly Arg Ser Cys Phe Thr Arg Arg Ile Glu Glu
 145 150 155 160
 Ala Arg Arg Ala Ser Gln Glu Pro Glu Gly Cys Ile Ala Leu Lys Gln
 165 170 175
 Ser Val Pro

<210> 31
 <211> 1226
 <212> DNA
 <213> Oryza sativa

<400> 31
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 cggagacctt ctccaggctc cgggagcagg gcaagactgc attcattcca ttcattaccg 300
 ccagtgaacc tgacttggca accacatcga aagcgttgaa gatccttgat tccgtgtggt 360
 cagatgtgat tgagctgggt gtaccttact cggatccgtt ggctgatggg ccggttattc 420
 aggcctgcagc aacgcgtgct cttaagaaag gcgctacatt tgattccgtc atagctatgc 480
 tcaagggggt tatacctgag ttgtcttgct caatagtatt cttcacatac tacaacccaa 540
 ttttaaagcg cggagtgaat aacttcatgg ctatcattaa acaagccggc gtacatgggtc 600
 ttgtagtacc tgatcttcct ttggaagaga cagcgtgttt gaggaatgag gctgtcatgc 660
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Lys Ser Leu Lys Ser Ala Leu Leu
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INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

| | | |
|--|-----------|---|
| (51) International Patent Classification ⁶ : C12N 15/60, 9/88, 1/21, C12Q 1/68 | A3 | (11) International Publication Number: WO 99/49058 (43) International Publication Date: 30 September 1999 (30.09.99) |
| (21) International Application Number: PCT/US99/06046 (22) International Filing Date: 19 March 1999 (19.03.99) (30) Priority Data: 60/079,386 26 March 1998 (26.03.98) US (71) Applicant (for all designated States except US): E.I. DU PONT DE NEMOURS AND COMPANY [US/US]; 1007 Market Street, Wilmington, DE 19898 (US). (72) Inventors; and (75) Inventors/Applicants (for US only): VOLLMER, Steven, J. [US/US]; 1019 Tweedbrook Road, Wilmington, DE 19810 (US). FALCO, Saverio, Carl [US/US]; 1902 Millers Road, Arden, DE 19810 (US). BROGLIE, Richard, M. [US/US]; 520 Port Royal Court, Landenberg, PA 19350 (US). BRYAN, Gregory, T. [NZ/US]; 1215 Spruce Street, Wilmington, DE 19805 (US). CAHOON, Rebecca, E. [US/US]; 2331 West 18th Street, Wilmington, DE 19806 (US). RAFALSKI, J., Antoni [US/US]; 2028 Longcome Drive, Wilmington, DE 19810 (US). (74) Agent: MAJARIAN, William, R.; E.I. du Pont de Nemours and Company, Legal Patent Records Center, 1007 Market Street, Wilmington, DE 19898 (US). | | (81) Designated States: AL, AU, BA, BB, BG, BR, CA, CN, CU, CZ, EE, GD, GE, HR, HU, ID, IL, IN, IS, JP, KP, KR, LC, LK, LR, LT, LV, MG, MK, MN, MX, NO, NZ, PL, RO, SG, SI, SK, SL, TR, TT, UA, US, UZ, VN, YU, ZA, ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG). Published <i>With international search report.</i> (88) Date of publication of the international search report: 6 April 2000 (06.04.00) |
| (54) Title: TRYPTOPHAN BIOSYNTHETIC ENZYMES (57) Abstract This invention relates to an isolated nucleic acid fragment encoding a tryptophan biosynthetic enzyme. The invention also relates to the construction of a chimeric gene encoding all or a portion of the tryptophan biosynthetic enzyme, in sense or antisense orientation, wherein expression of the chimeric gene results in production of altered levels of the tryptophan biosynthetic enzyme in a transformed host cell. | | |

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| DK | Denmark | LR | Liberia | SG | Singapore | | |
| EE | Estonia | | | | | | |

INTERNATIONAL SEARCH REPORT

International Application No.

PCT/US 99/06046

A. CLASSIFICATION OF SUBJECT MATTER

IPC 6 C12N15/60 C12N9/88 C12N1/21 C12Q1/68

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C12N C12Q

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

| Category | Citation of document, with indication, where appropriate, of the relevant passages | Relevant to claim No. |
|----------|---|-----------------------|
| X | WO 97 26366 A (DEKALB GENETICS CORP ;ANDERSON PAUL C (US); CHOMET PAUL S (US); GR) 24 July 1997 (1997-07-24) the whole document --- -/-- | 1-34, 54-59 |

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

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 "Z" document member of the same patent family

Date of the actual completion of the international search

18 January 2000

Date of mailing of the international search report

03.02.00

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 European Patent Office, P.B. 5818 Patentlaan 2
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 Fax: (+31-70) 340-3016

Authorized officer

Smalt, R

INTERNATIONAL SEARCH REPORT

International Application No
PCT/US 99/06046

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

| Category | Citation of document, with indication where appropriate of the relevant passages | Relevant to claim No |
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| | | |
|---|---|-------------------------------------|
| X | <p>DATABASE EMBL - EMBEST20 Online! Entry ZM248. Acc.no.T25248. 1 October 1994 (1994-10-01) SHEN, B. ET AL.: "5c10h08 membrane-free polysomes from endosperm Zea mays cDNA clone 5c10h08 5' end similar to anthranilate synthase." XP002112345 the whole document -& SHEN, B. ET AL.: "Partial sequencing and mapping of clones from two maize cDNA libraries." PLANT MOLECULAR BIOLOGY, vol. 26, 1994, pages 1085-1101, XP002042536 the whole document</p> | <p>1,2,10, 18,22, 23,28</p> |
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INTERNATIONAL SEARCH REPORT

International Application No

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INTERNATIONAL SEARCH REPORT

International Application No
PCT/US 99/06046

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| P,X | WO 99 11800 A (HASEGAWA HISAKAZU ;HOKKO CHEM IND CO (JP); TERAKAWA TERUHIKO (JP);) 11 March 1999 (1999-03-11) page 125 -page 131; claims 1,2 -& DATABASE WPI Section Ch, Week 9919 Derwent Publications Ltd., London, GB; Class C06, AN 99-228982 XP002112346 the whole document | 1-13, 18-21, 54-58 |

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US 99/06046

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:
2. ☒ Claims Nos.: 57, 58
because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
see FURTHER INFORMATION sheet PCT/ISA/210
3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

see additional sheet

1. ☒ As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
- ☒ No protest accompanied the payment of additional search fees.

INTERNATIONAL SEARCH REPORT

International Application No. PCT/US 99/06046

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

Continuation of Box I.2

Claims Nos.: 57,58

Present claims 57 and 58, relating to products which can be obtained by the methods of claims 55 and 56, respectively, have not been searched due to lack of support for such products within the meaning of Article 6 PCT and/or lack of disclosure of such products within the meaning of Article 5 PCT in the description.

The applicant's attention is drawn to the fact that claims, or parts of claims, relating to inventions in respect of which no international search report has been established need not be the subject of an international preliminary examination (Rule 66.1(e) PCT). The applicant is advised that the EPO policy when acting as an International Preliminary Examining Authority is normally not to carry out a preliminary examination on matter which has not been searched. This is the case irrespective of whether or not the claims are amended following receipt of the search report or during any Chapter II procedure.

INTERNATIONAL SEARCH REPORT

International Application No. PCT/US 99/06046

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

This International Searching Authority found multiple (groups of) inventions in this international application, as follows:

1. Claims: 2,3,10,11,23,24,27,28 completely, and 1,18-22, 31-34,54-59 partially

Corn anthranilate synthase subunits alpha and beta, portions thereof, nucleic acids encoding them and sequences with at least 80% homology thereto, chimeric genes comprising a portion of said nucleic acids, hosts transformed with said nucleic acids, method for altering expression of said gene, method for obtaining said nucleic acids, and method for evaluating the ability of a compound to inhibit the activity of said protein.

2. Claims: 36,37,43,44 completely, and 35,50-59 partially

As invention 1, but limited to corn tryptophan synthase subunit alpha.

3. Claims: 4,12,25,29 completely, and 1,18-22, 31-34, 54-59 partially

As invention 1, but limited to rice anthranilate synthase subunits alpha and beta.

4. Claims: 38,39,45,46 completely, and 35,50-59 partially

As invention 1, but limited to rice tryptophan synthase subunit alpha.

5. Claims: 5,6,13,14,40,47 completely, and 1,18-21,35, 50-59 partially

As invention 1, but limited to soybean anthranilate synthase subunit alpha and tryptophane synthase subunit alpha.

6. Claims: 7-9,15-17,26,30,41,42,48,49 completely, and 1, 18-22,31-35,50-59 partially

As invention 1, but limited to wheat anthranilate synthase subunits alpha and beta, and tryptophane synthase subunit alpha.

INTERNATIONAL SEARCH REPORT

information on patent family members

International Application No

PCT/US 99/06046

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